

Engineered neuronal assemblies and functional connectivity analysis

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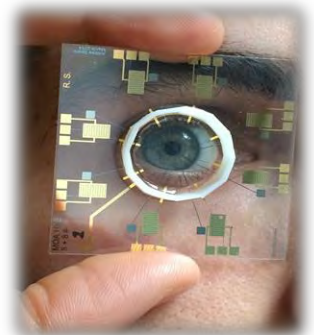
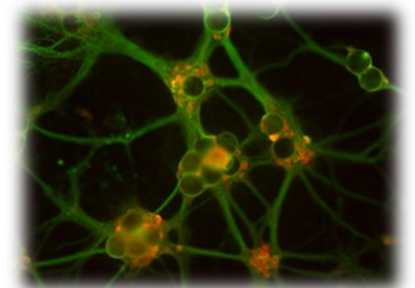
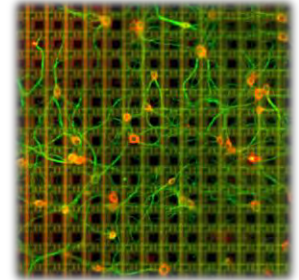
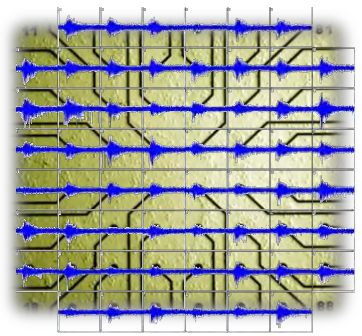
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Dibris

Outline

- CMOS based MEAs
- Neural signal analysis and connectivity
- Neural activity modulation by nanoparticles



Neural-interfaces: technological advances

“(...) Progress in **large-scale recording** of neuronal activity depends on the development of three critical components: **the neuron-electrode interface**, **methods for spike sorting /identification and tools for the analysis and interpretation of parallel spike trains**. In addition to **increasing the numbers of recording sites on silicon probes**, the development of on-chip interface circuitry is another priority. (...)”

from G. Buzsáki, “ Large-scale recording of neuronal ensembles”, Nature Neuroscience, Vol. 7, No. 7, May 2004

- large under-sampling of the network activity (~10'000:100)
- limited number of microelectrodes (60-120)
- limited electrode pitch (~100 μm)



need of new enabling technologies
need of new analysis methods

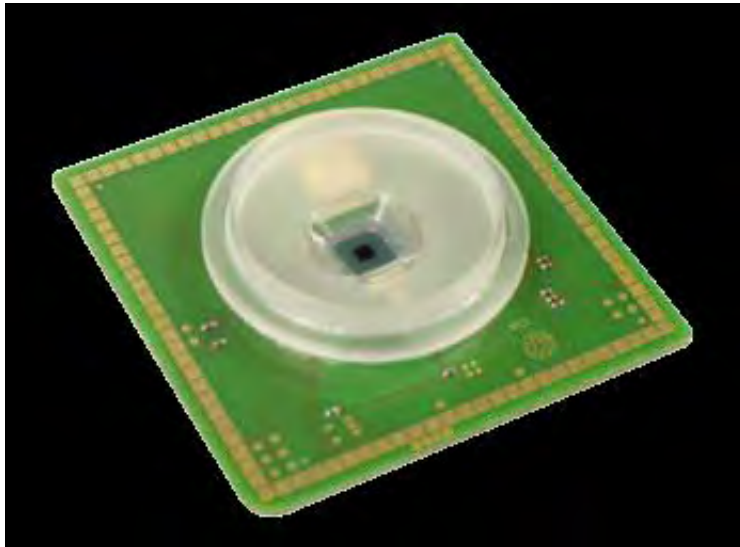
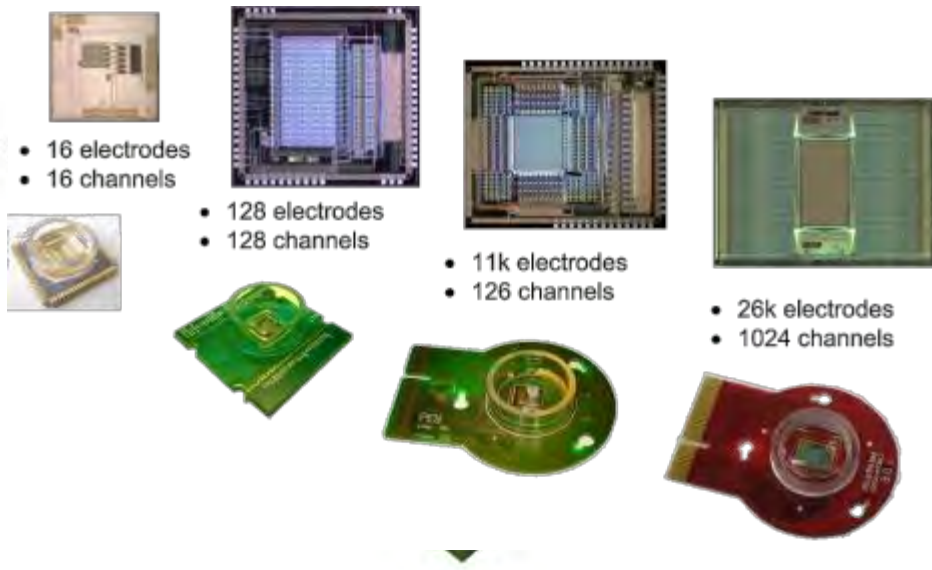
CMOS based approach

Hierleman group ETH

- CMOS-based microelectrode array with 11'016 metal electrodes
- 128 addressable electrodes at a sampling rate of 20kHz
- Now improved version

Herr et al., Biosensors and Bioelectronics, (2007), pp. 2546–2553

maxwell
BIOSYSTEMS



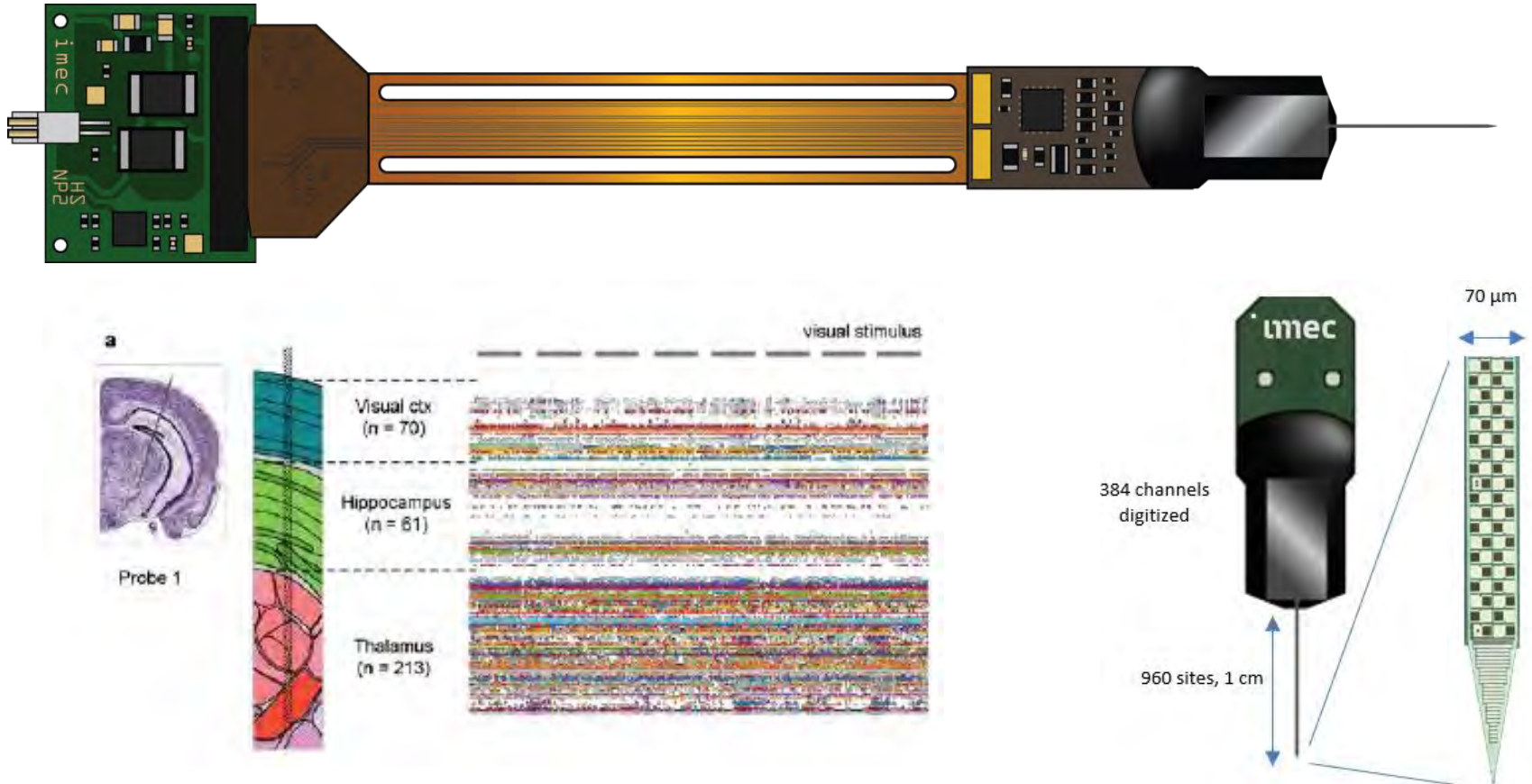
- 4225 Field Effect Transistor array with 1024 stimulating sites
- recording from all electrodes at a sampling rate of 25 kHz
- High-signal quality

multichannel*
systems

CMOS based approach for in vivo

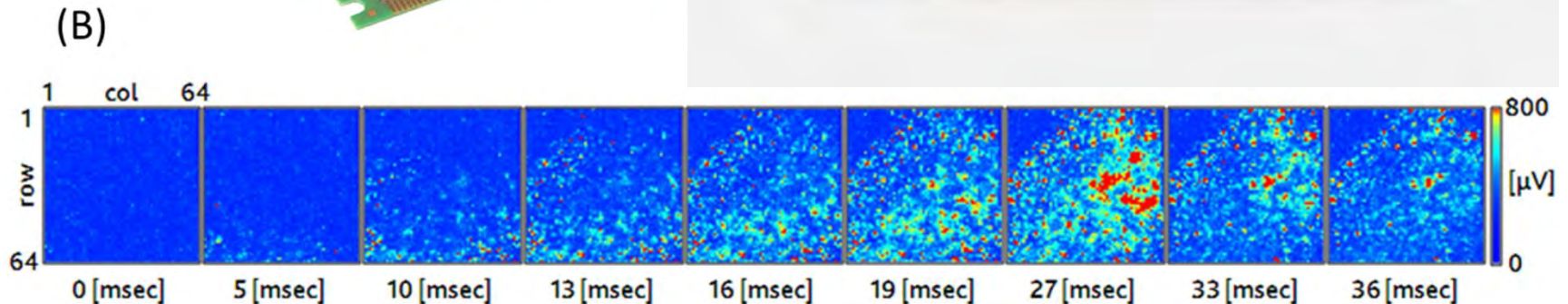
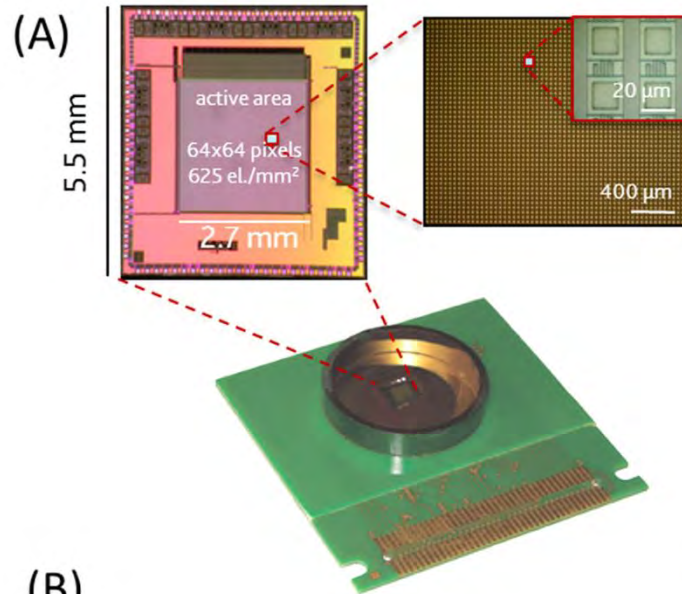
Nature (2017)

Fully Integrated Silicon Probes for High-Density Recording of Neural Activity




T. Harris et al., Janelia Research Campus

High-density CMOS based device



- IDEA: UE NEST project (2005-2008)
- Start-up (in Switzerland) (2011- www.3brain.com)



Samlab's  ÉCOLE POLYTECHNIQUE
FÉDÉRALE DE LAUSANNE
Sensors, Actuators and
Microsystems Lab

A long-story: 15 years of experience in CMOS-MEAs



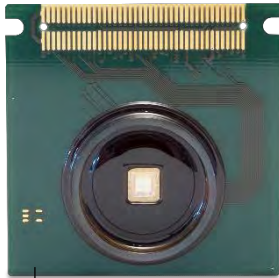
High-density large scale CMOS-MEAs

APOLLO
Gen 0-1

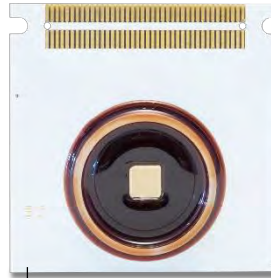


4096 recording el.
42 μm pitch
2.7 x 2.7 mm² sensing area
3 x 3 mm² flat area

ARTEMIS
Gen 2

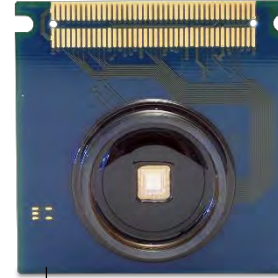


4096 recording el.
42 μm pitch
2.7 x 2.7 mm² sensing area
6 x 6 mm² flat area



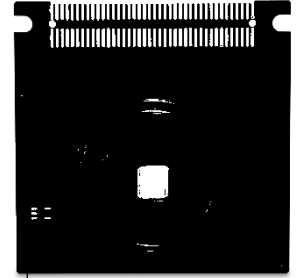
4096 recording el.
16 stim el.
81 μm pitch
5.1 x 5.1 mm² sensing area
6 x 6 mm² flat area

KHIRON
Gen 3



4096 recording el.
4096 stim el.
60 μm pitch
3.8 x 3.8 mm² sensing area
6 x 6 mm² flat area

TBD
Gen 4



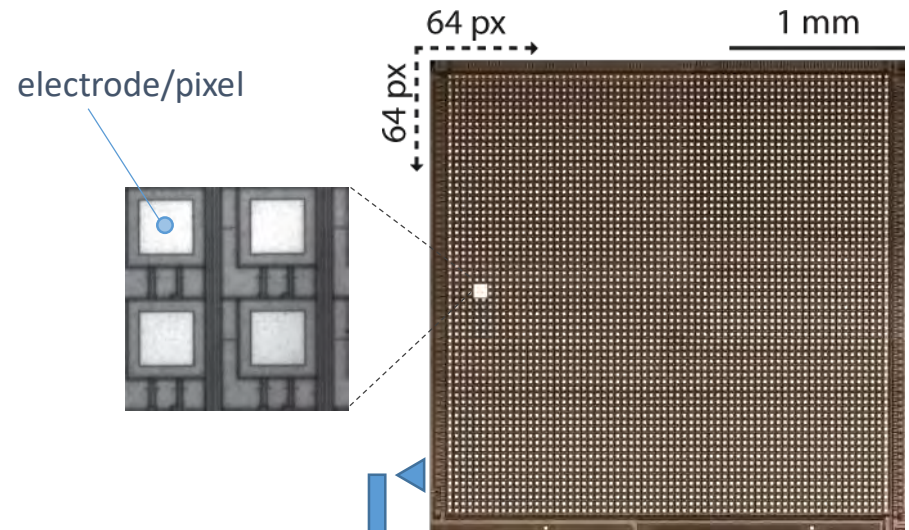
Under development

High-density CMOS based device

The Active Pixel Sensor (APS) technology

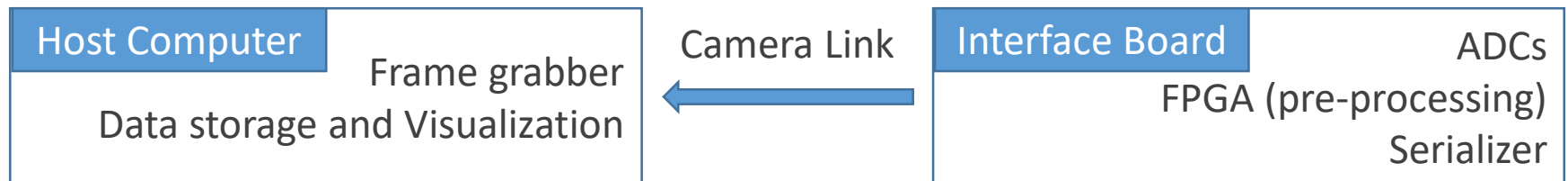
- redesigned in order to sense the electrophysiological signals
- each electrode/pixel integrates a pre-amplifier

no. of electrodes	4096
electrode size	21 μm
electrode separation	21 μm
active area	$\sim 7 \text{ mm}^2$
spatial density	$\sim 580 \text{ el/mm}^2$
sampling rate	7.7 - 125 kHz
data rate	$\sim 0.5 \text{ Gbit/s}$

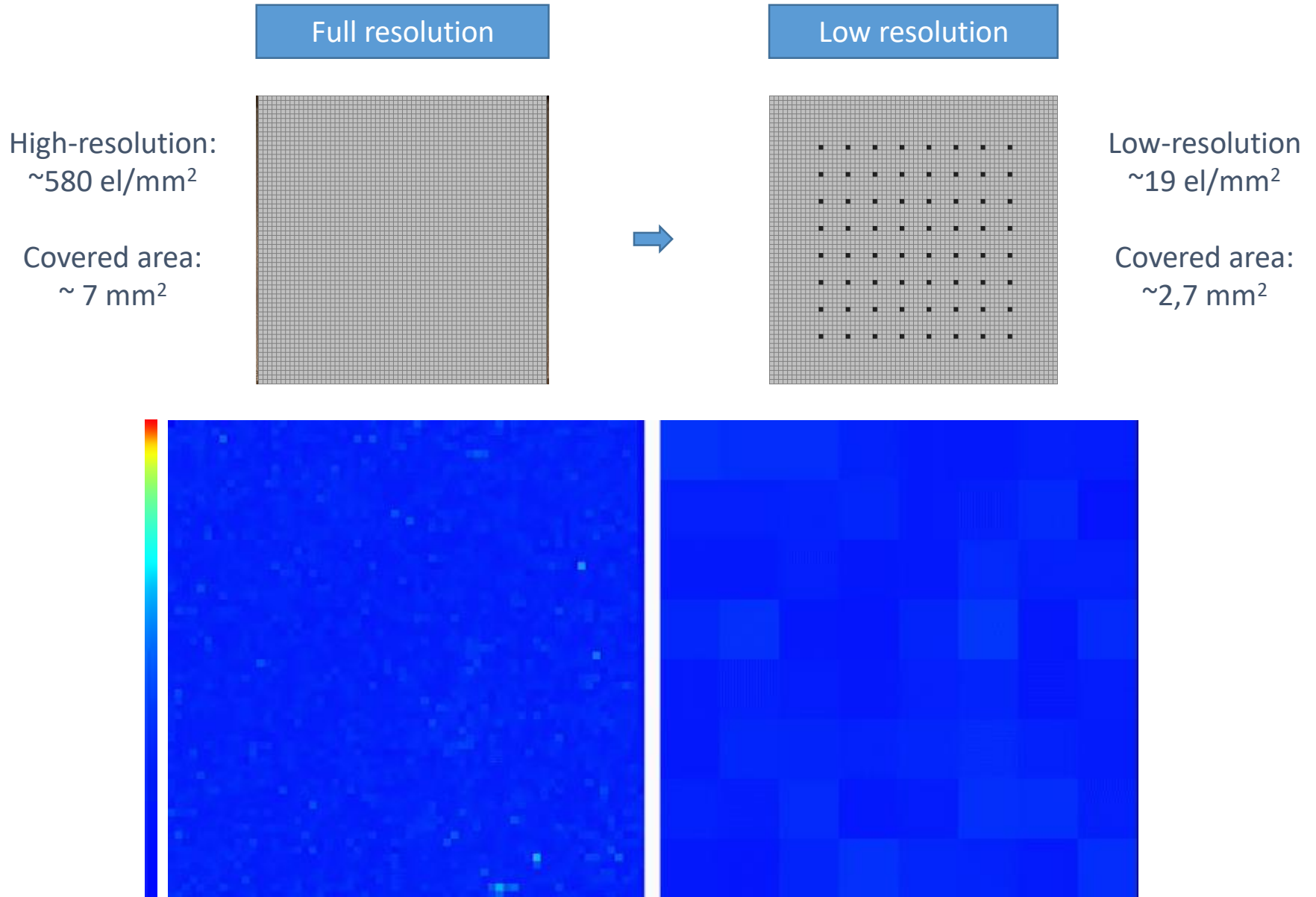


The high-density APS-MEA platform

- oriented to image/video concepts



2D networks on high-density APS



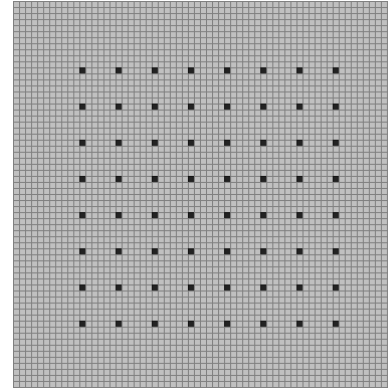
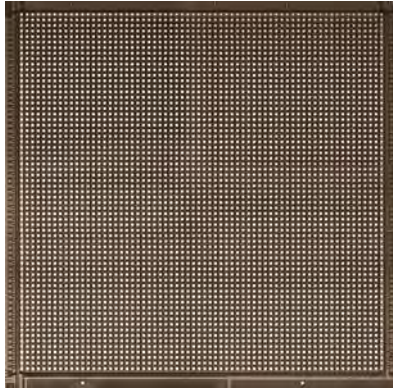
2D networks on high-density APS

Full resolution

Low resolution

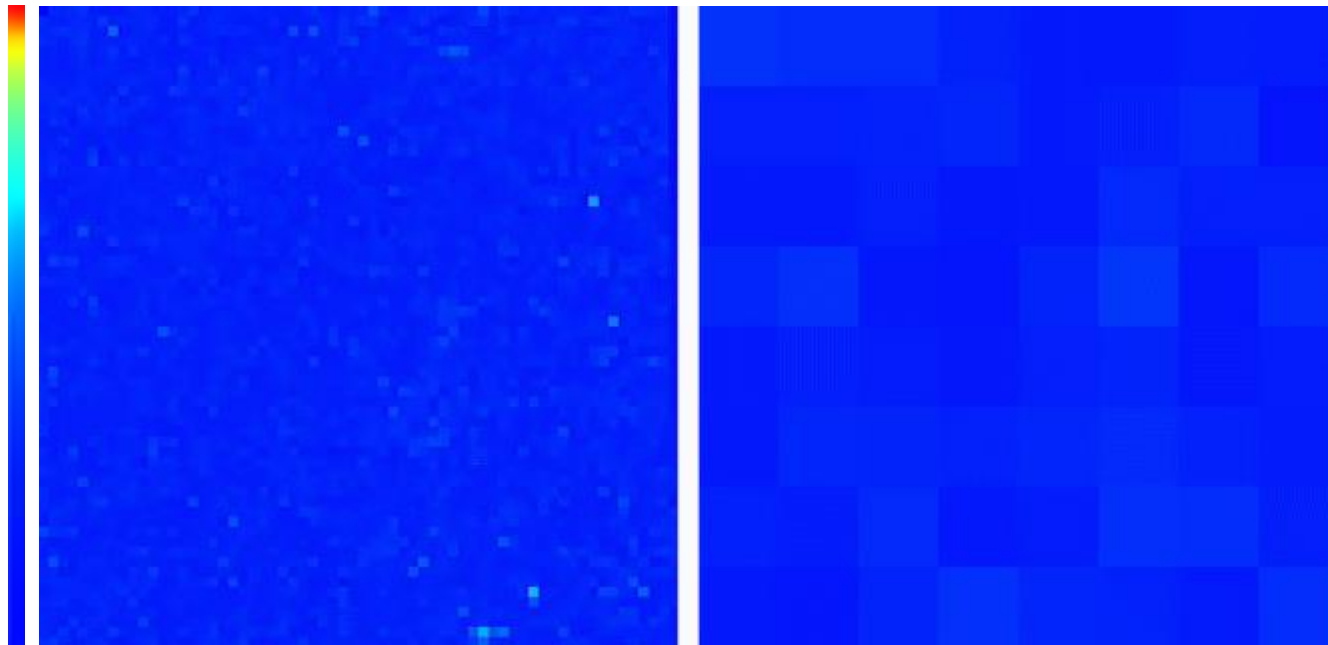
High-resolution
(~ 580 el/mm²)

Large-scale
(~ 7 mm²)

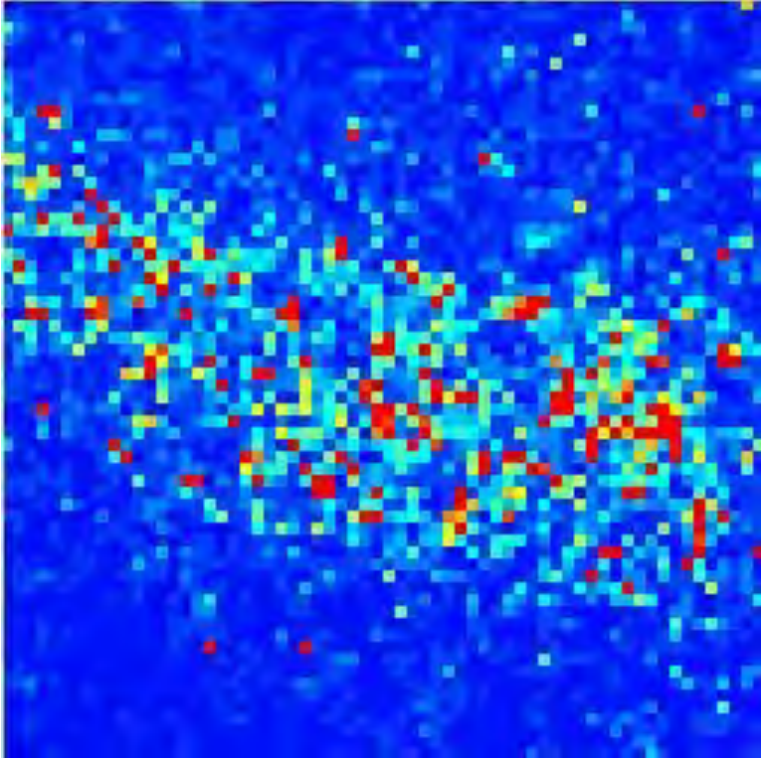


Low-resolution
(~ 19 el/mm²)

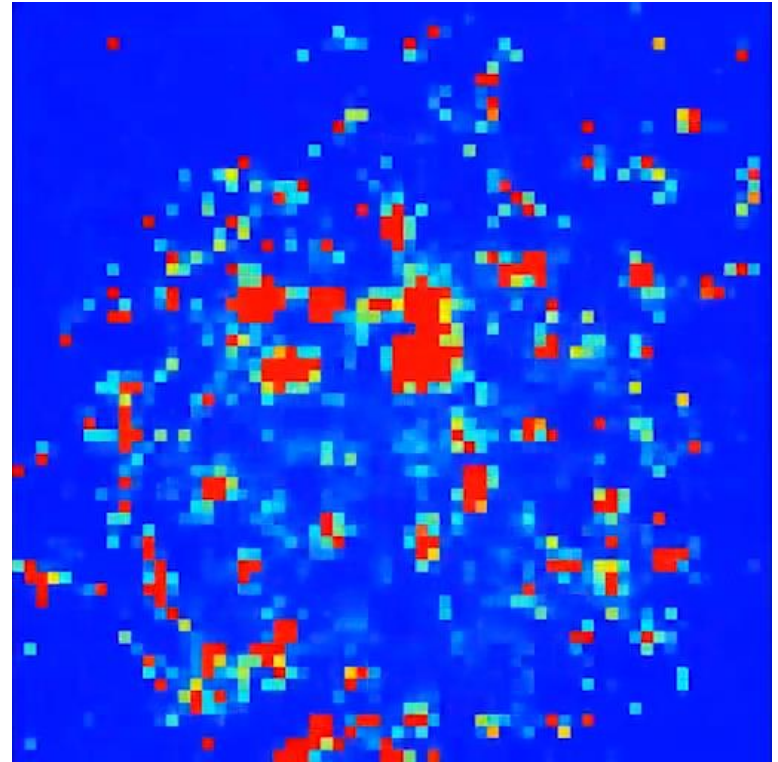
Medium-scale
($\sim 2,7$ mm²)



Example: dissociated cultures – whole network synchronous activity

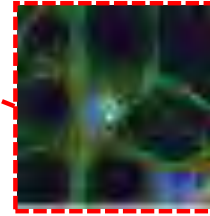
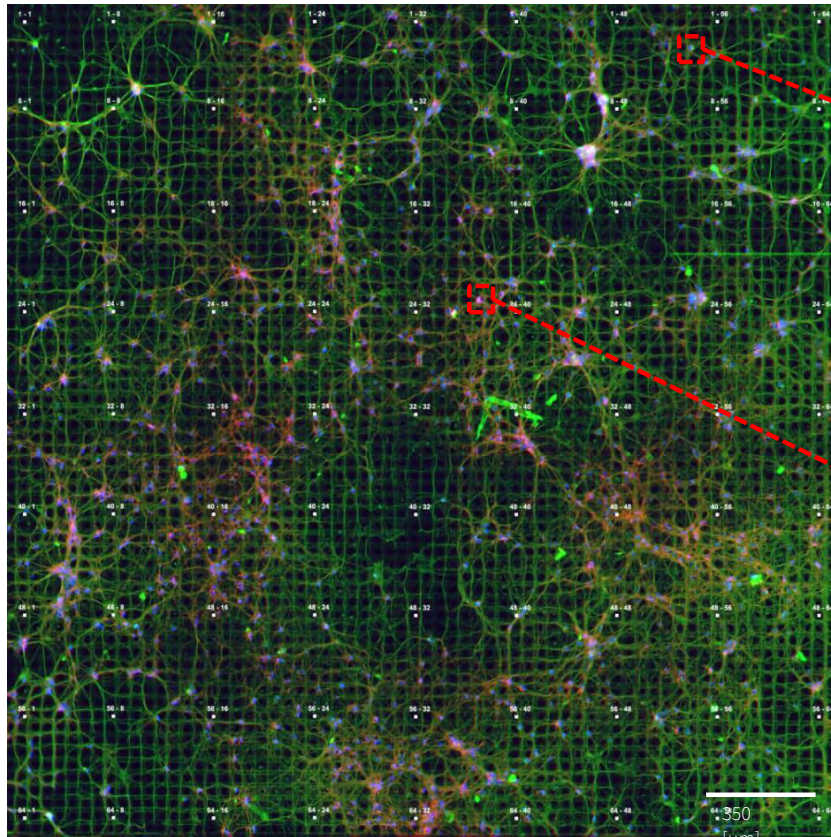


Post natal 14 DIVs mouse culture
30 msec synchronous event

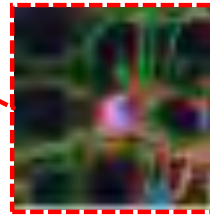


Embryonic 22 DIVs rat culture
100 msec synchronous event

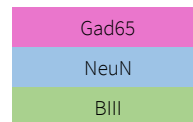
Example: structural and functional identification of sub-networks



40
[μm]
excitatory not GABAergic neuron

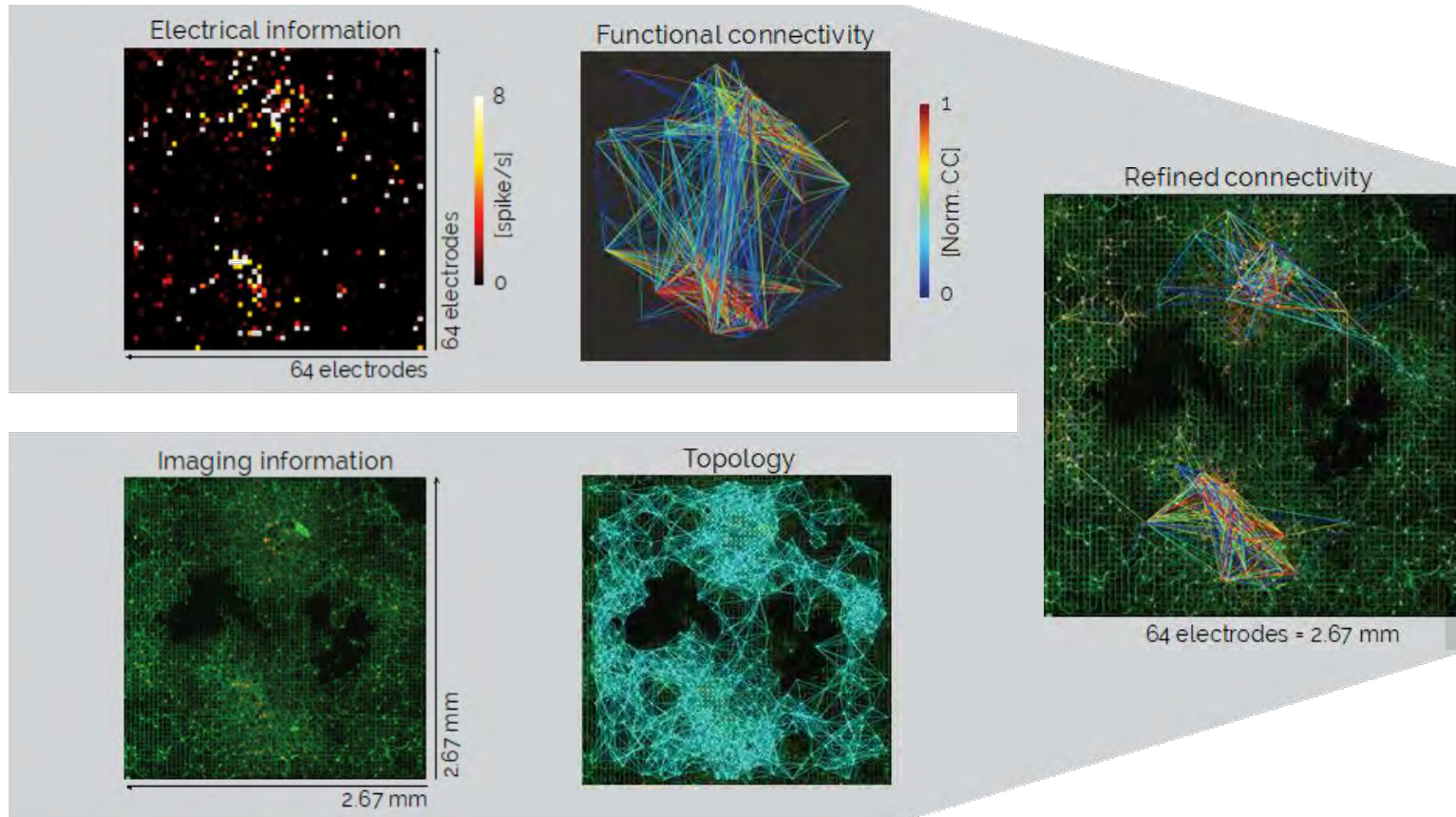


40
[μm]
inhibitory GABAergic neuron

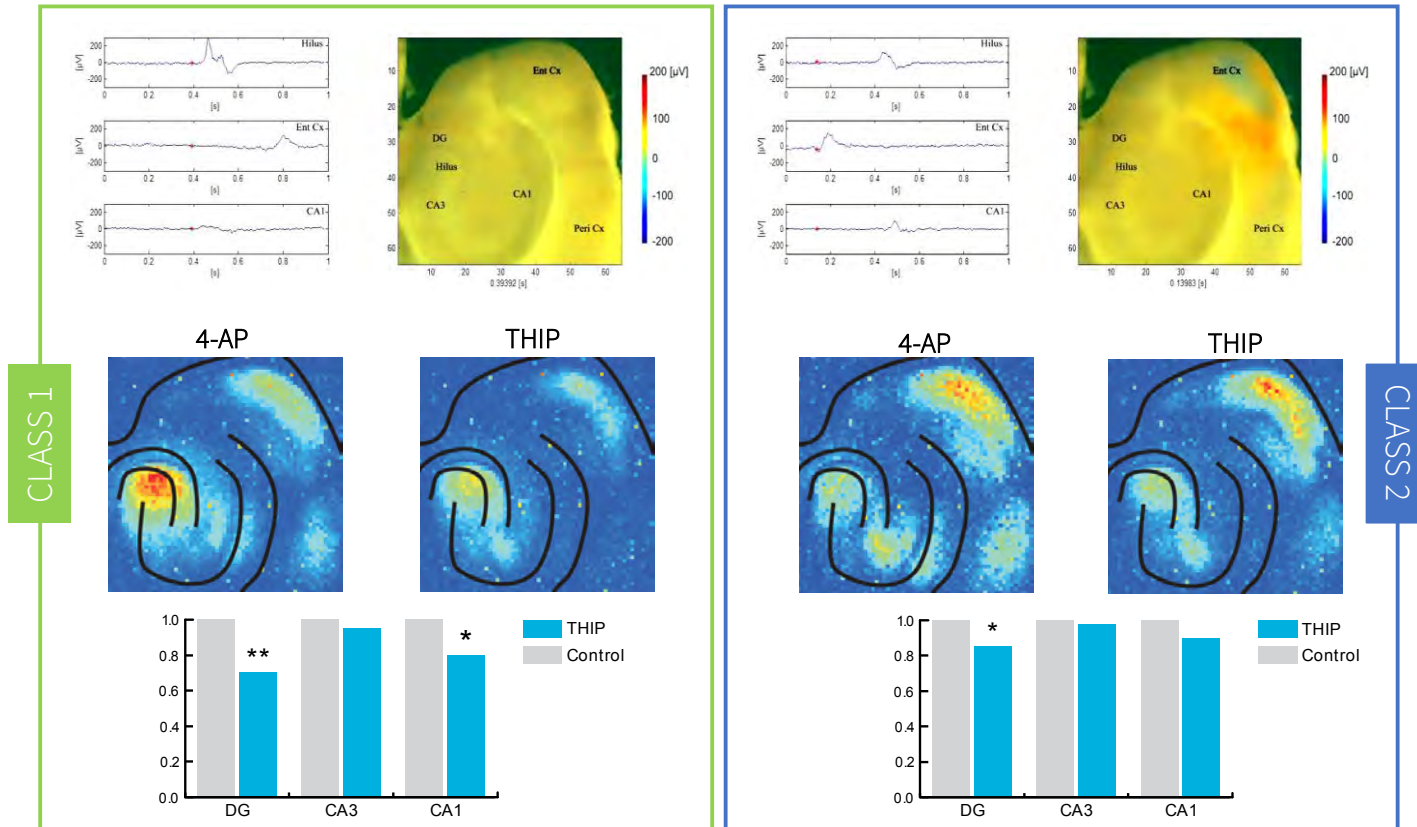


	Basal		Bic 30 μM		TTEST
MFR	mean	stdErr	mean	stdErr	
Exc	0.63	0.05	0.88	0.08	**
Inh	1.14	0.08	1.36	0.13	-
TTEST	**		*		

Example: coupling electrophysiological and topological info

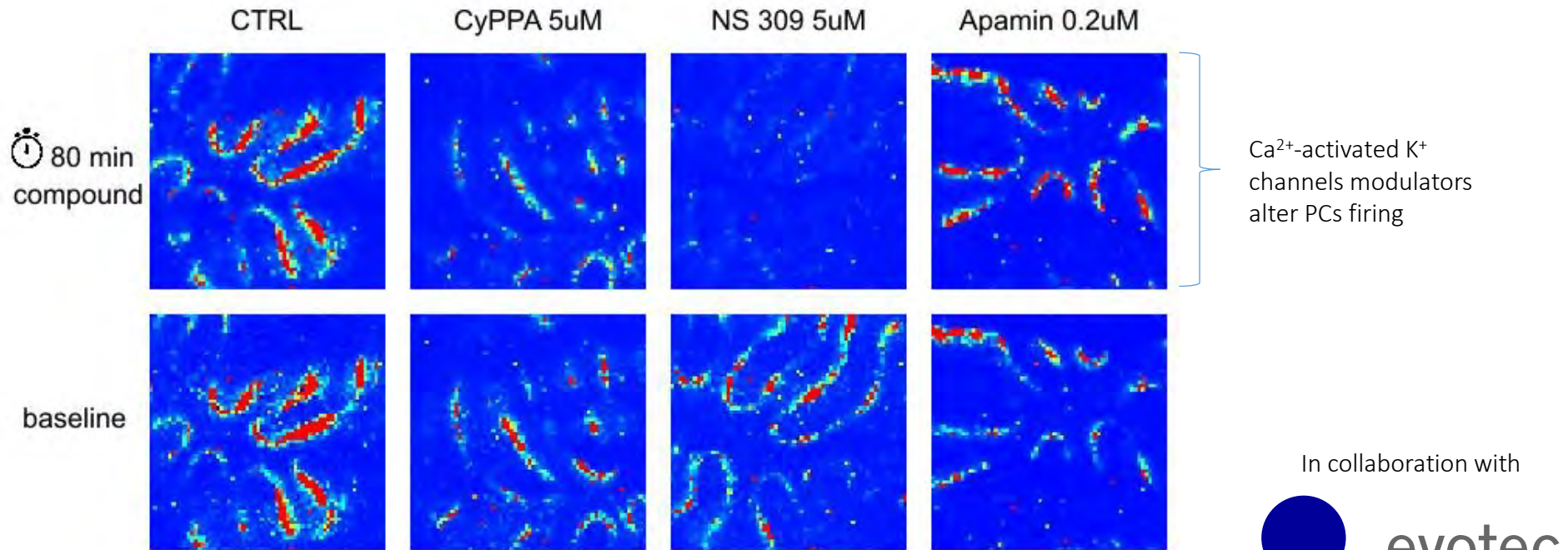


Example: epileptic model for anticonvulsant compound testing



Ferrea et al. 2012, Front. Neural Circuits

Example: compound effect on purkinje activity in cerebellum slice



A Ugolini et al. – Fens 2018

Engineered neuronal assemblies: data analysis

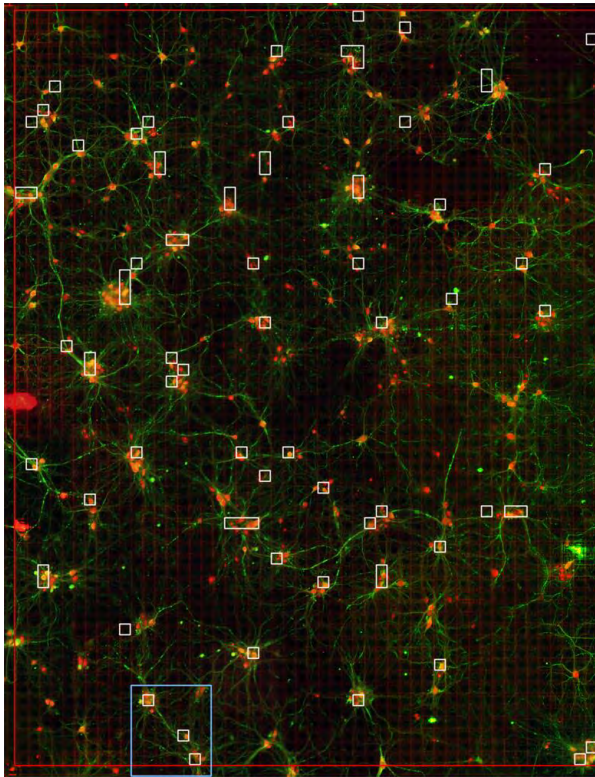
“(...) Progress in **large-scale recording** of neuronal activity depends on the development of three critical components: **the neuron-electrode interface**, **methods for spike sorting /identification and tools for the analysis and interpretation of parallel spike trains**. In addition to **increasing the numbers of recording sites on silicon probes**, the development of on-chip interface circuitry is another priority. (...)”

from G. Buzsáki, “ Large-scale recording of neuronal ensembles”, Nature Neuroscience, Vol. 7, No. 7, May 2004



need of new enabling technologies
need of new analysis methods

Structural vs functional connectivity

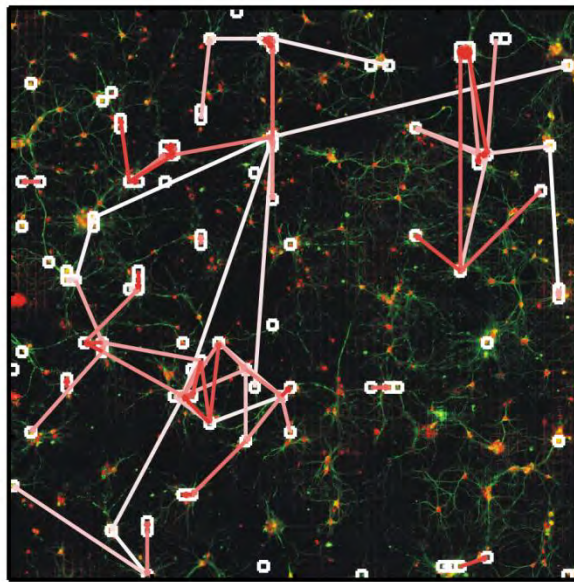


Functional connectivity
estimated by means of Cross-
Correlation based techniques
and information theory methods

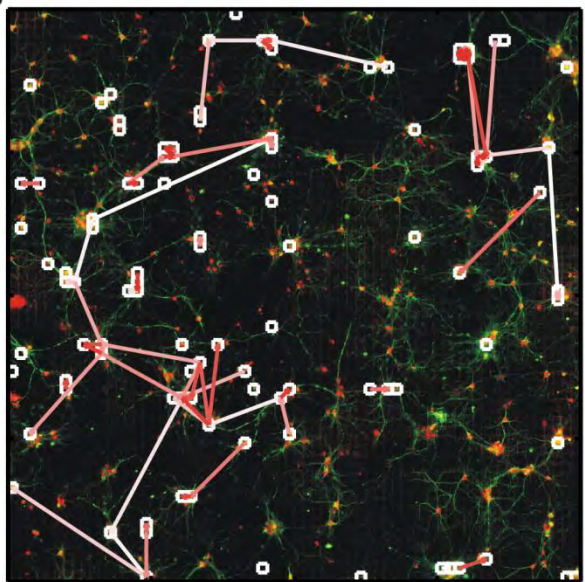
Garofalo et al. Plos One, (2009)

Maccione et al. J. Neurosci Methods (2012)

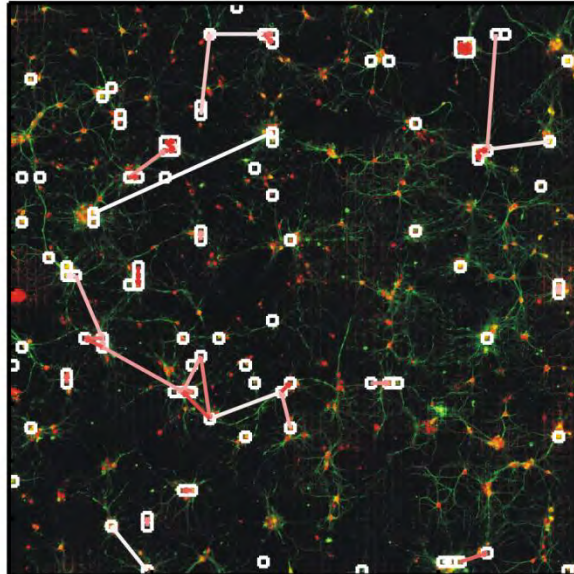
A



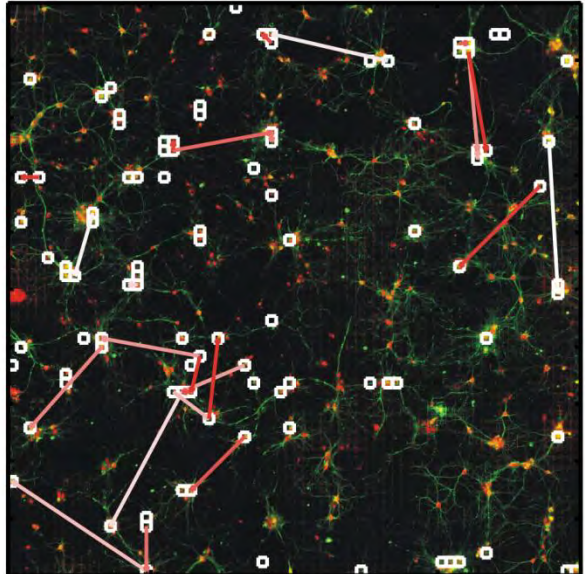
B



C



D



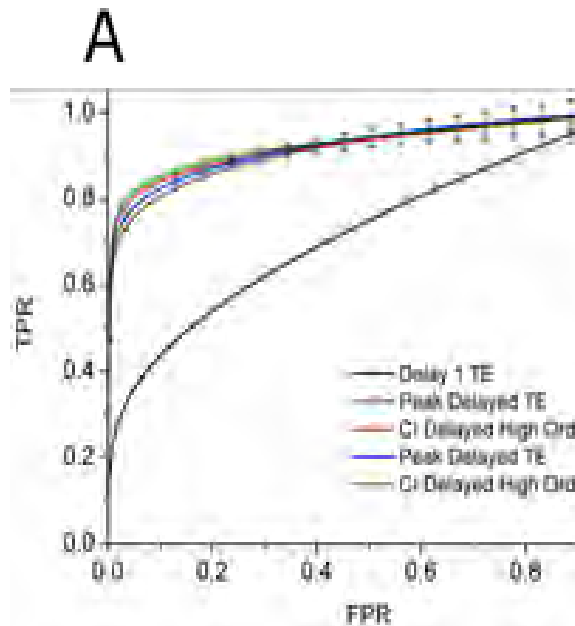
Functional-effective connectivity methods: Transfer Entropy revisited



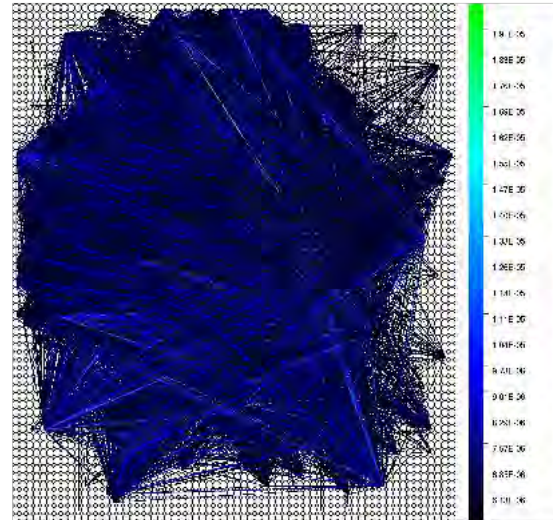
Vito Paolo Pastore, PhD student

$$TE_{y \rightarrow x} = \sum_{x_t x_{t-1} y_{t-1}}$$

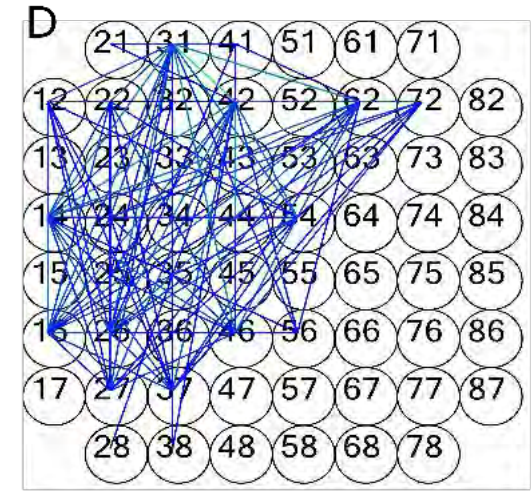
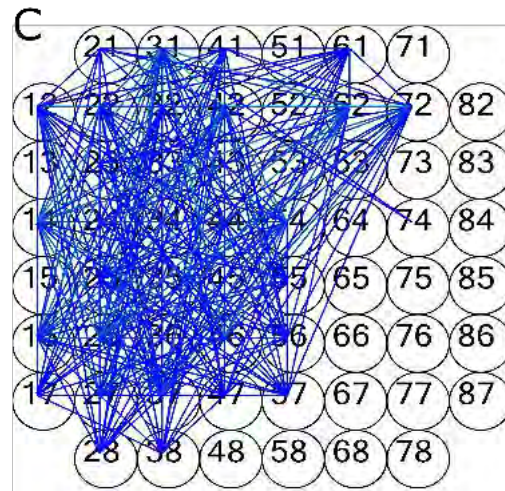
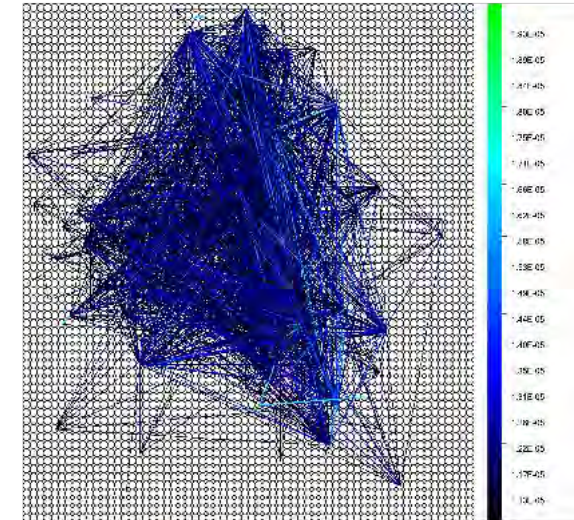
- for a reference spike train
- the couple (k, l) defines



A



B



Simple cross-correlation revisited

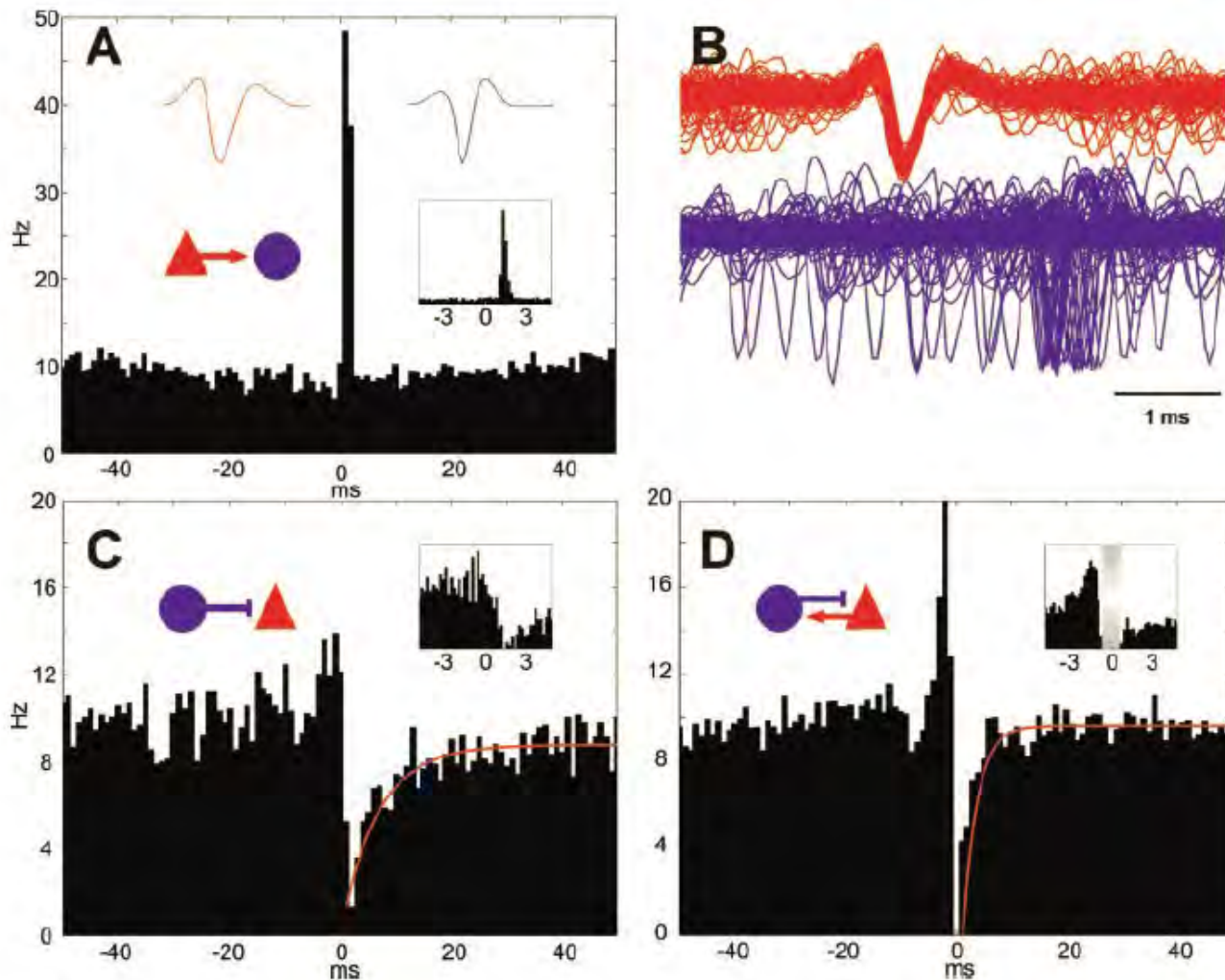
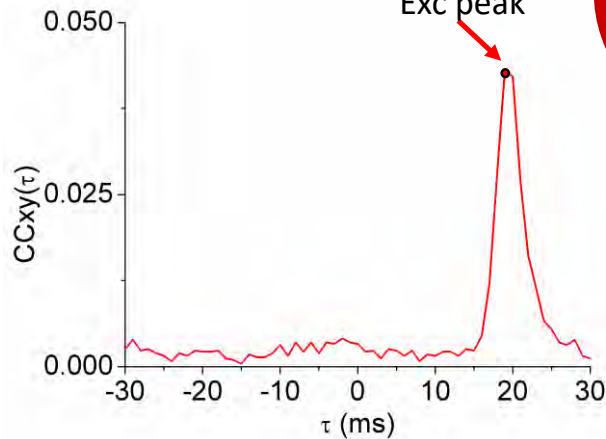


FIG. 2. Short-latency, monosynaptic interactions between neuron pairs. *A*: excitatory drive by a putative pyramidal cell (red triangle). Note large, sharp peak at ~ 2 ms in the cross-correlogram. Reference event is the spike of the putative pyramidal neuron (*time 0*). *Inset*: higher temporal resolution of the histogram. Averaged waveforms of the units (filtered: 600 Hz to 5 kHz) are also shown. On the bases of spike duration, the target cell was classified as a putative interneuron (blue circle; see text). *B*: superimposed traces of the neuron pair from 2 recording sites with the largest amplitude for each spike. Arrow, monosynaptically driven spikes. *C*: inhibitory suppression. Reference event: spike of the putative interneuron (blue circle). Note strong and immediate suppression of target spikes. The 2 neurons were recorded from different shanks (200- μ m lateral separation). Red line indicates exponential fit of suppression time course. *D*: reciprocal monosynaptic interactions of neurons recorded from the same shank. Reference event: spike of the putative interneuron (blue circle). Note excitation of the putative interneuron and strong suppression of the pyramidal cell (red triangle) spikes by the interneuron. Shading indicates the blank period of spike sampling (see METHODS).

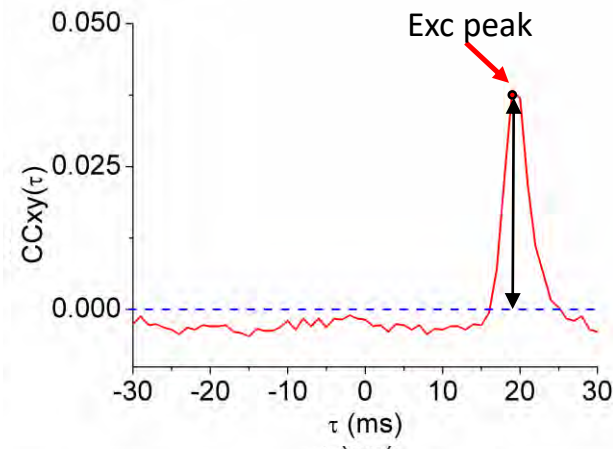
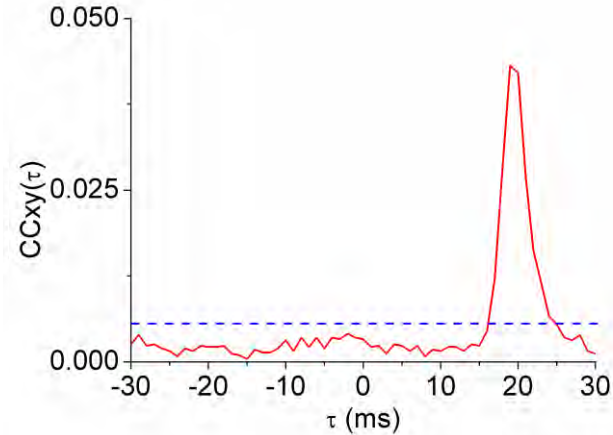
Functional-effective connectivity methods: Cross-Correlation revisited

$$C_{xy}(\tau) = \frac{1}{\sqrt{N_x N_y}} \sum_{s=1}^{N_x} \sum_{ti=(\tau-\nabla\tau/2)}^{(\tau+\nabla\tau/2)} x(t_s) y(t_s - t_i)$$

CC is able to detect the inhibitory links
Exc peak

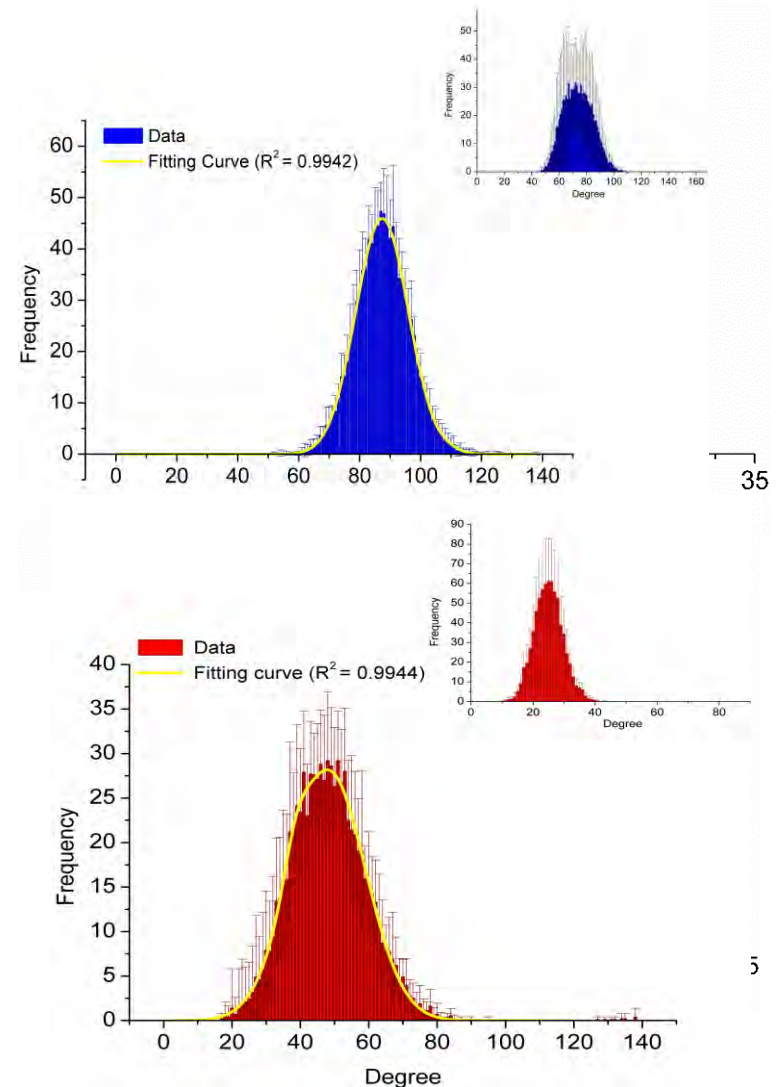


$$CC_{xy_peak_value} = \max_{\tau = [-W/2, +W/2]} \left\{ C_{xy}(\tau) - \frac{1}{W} \sum_{\tau=-W/2}^{\tau=+W/2} C_{xy}(\tau) \right\}$$



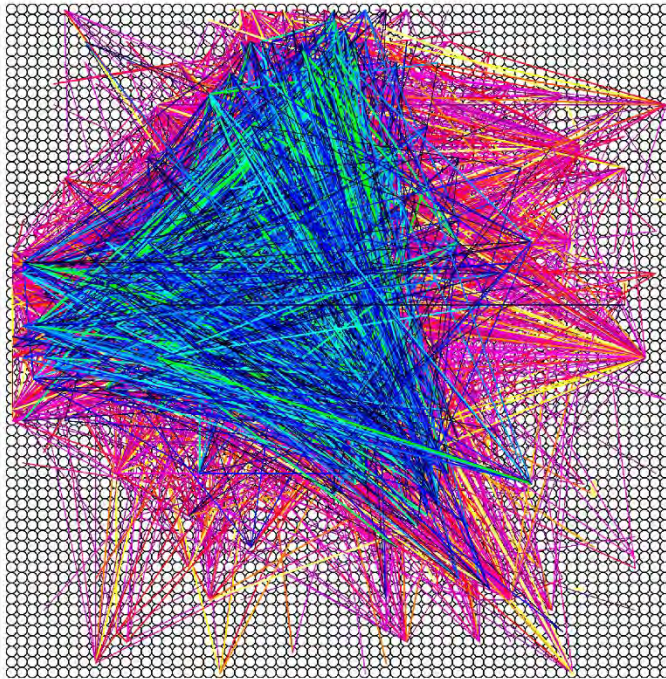
Filtered Normalized Cross-Correlation Histogram (FNCCH)

- It has been validated on in silico neural networks with **1000** neurons
- Identification of inhibitory links!
- Improvement of excitatory link detection
- Very good delay reconstruction
- Very good degree distribution reconstruction



Functional-effective connectivity

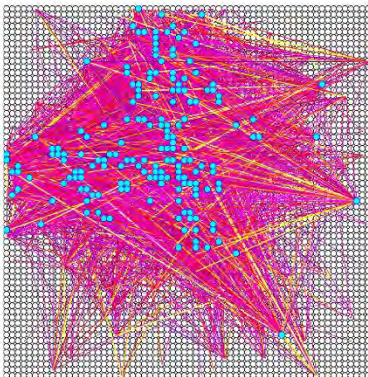
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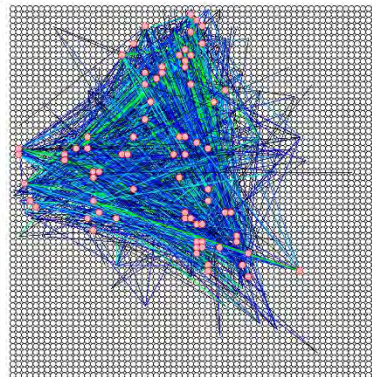
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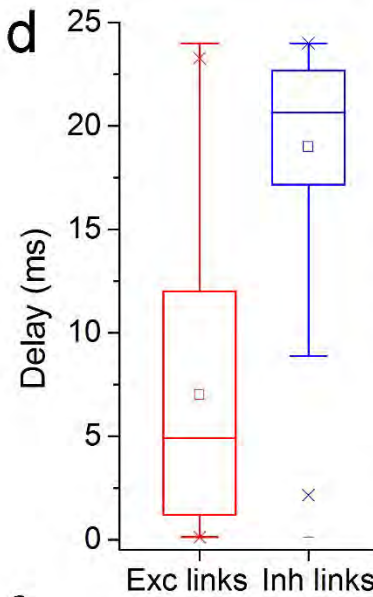
g b



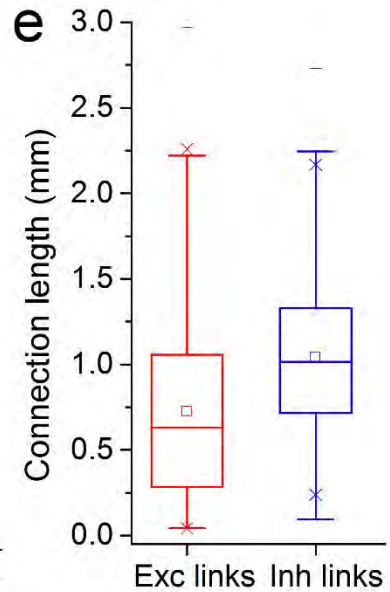
c



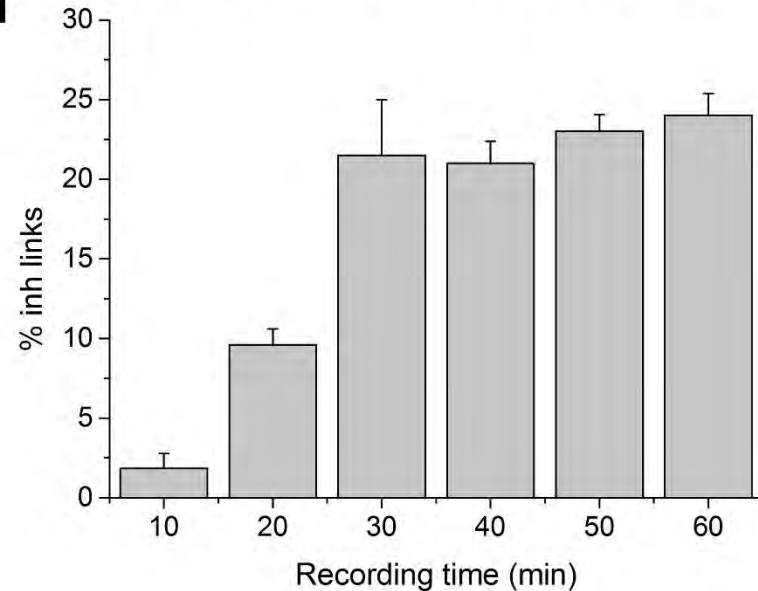
d



e



f

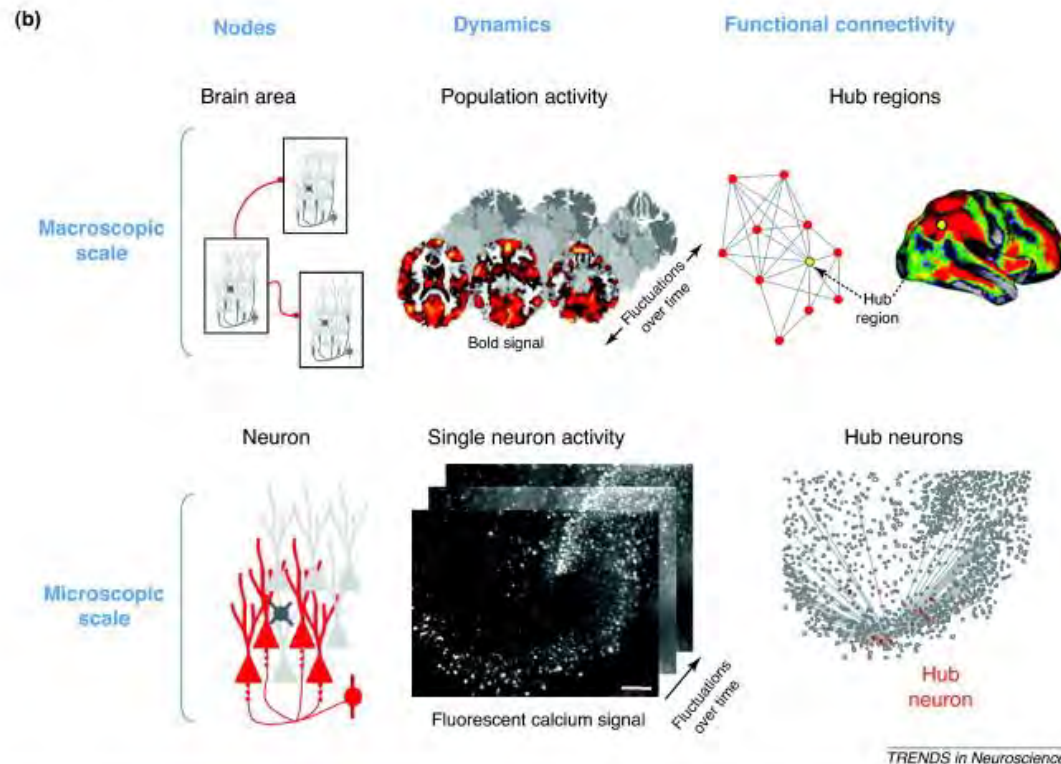
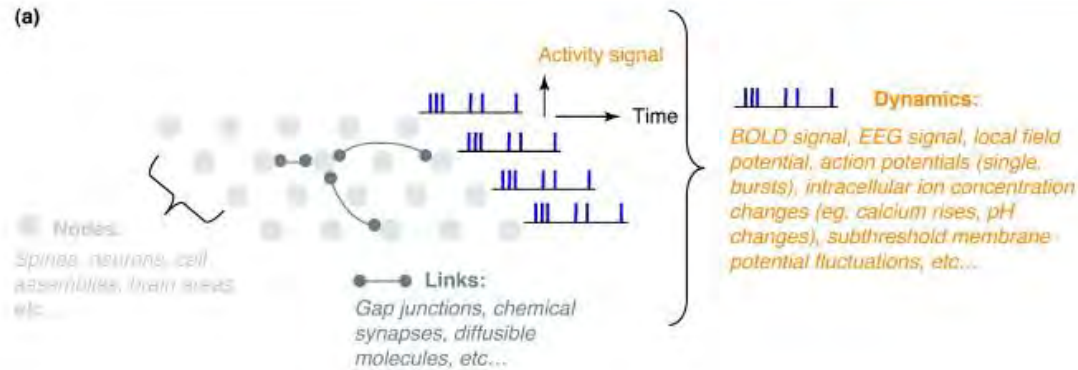


Connectivity & dynamics

Graph Theory can be used to:

explore and compare structural and functional brain networks

classify => topology



Feldst, S., Bonifazi P., Cossart R., "Dissecting functional connectivity of neuronal microcircuits: experimental and theoretical insight". *TINS*, 34, 225, (2011)

Graph theory and connectivity

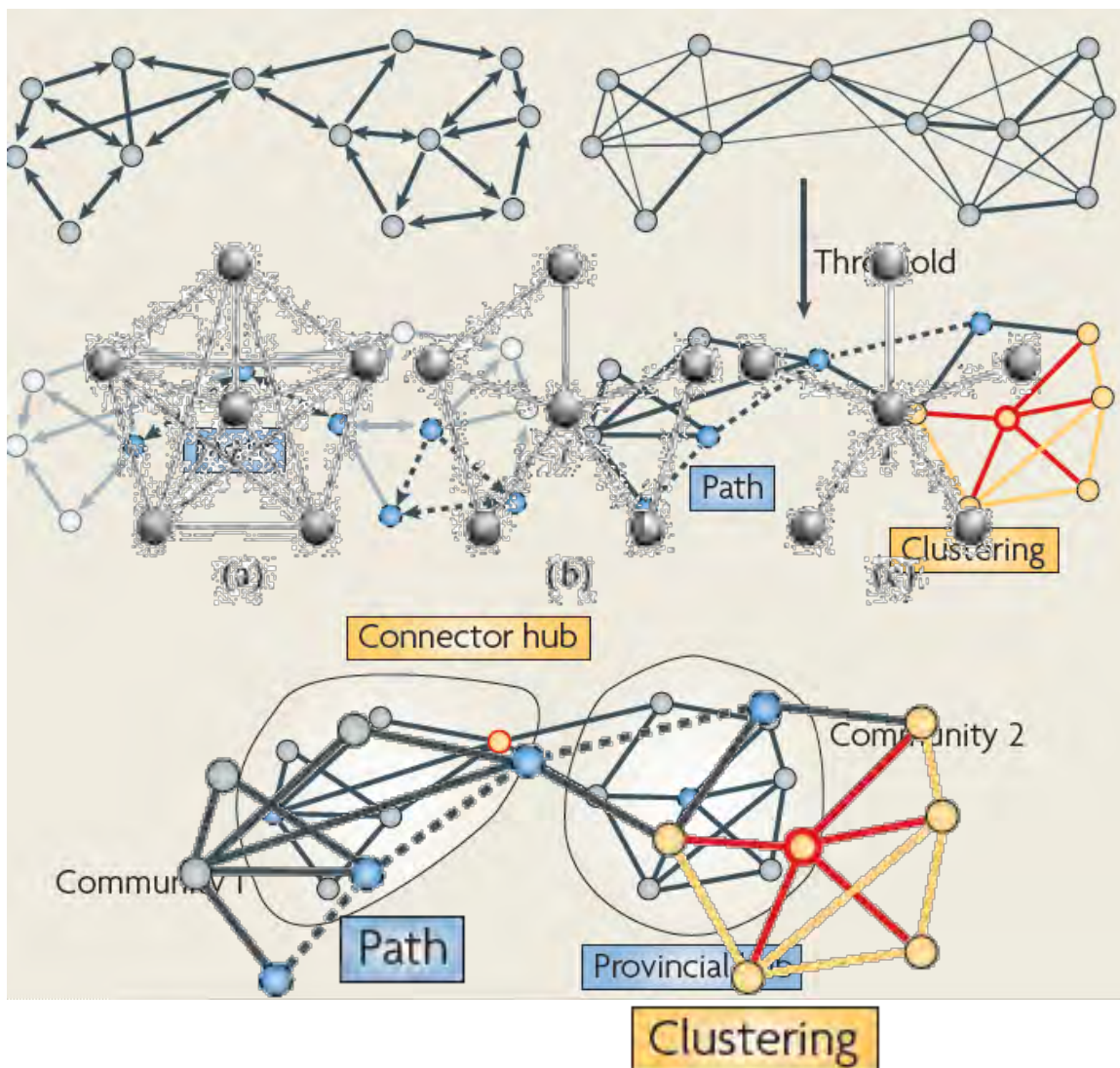
Topology of the network



parameters

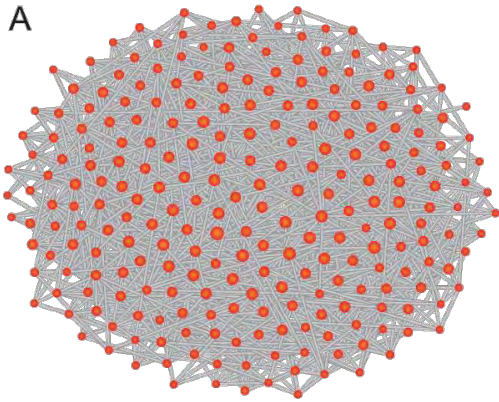
Clustering Coefficient (CC):
quantifies the number of connections that exist between the nearest neighbours of a node.

Mean Path Length (PL):
minimum number of edges that must be traversed to go from one node to another.



Network models

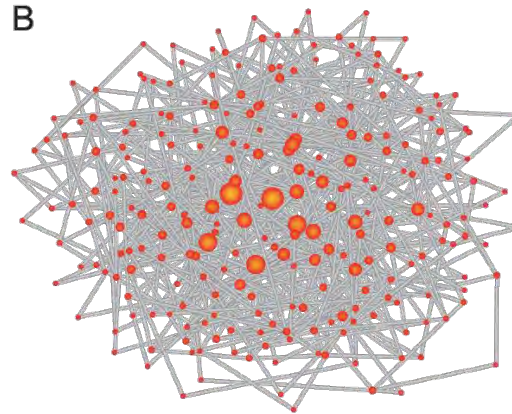
Random (RND) networks



Each pair of nodes has an equal probability of being connected

Degree distribution follows a Gaussian distribution

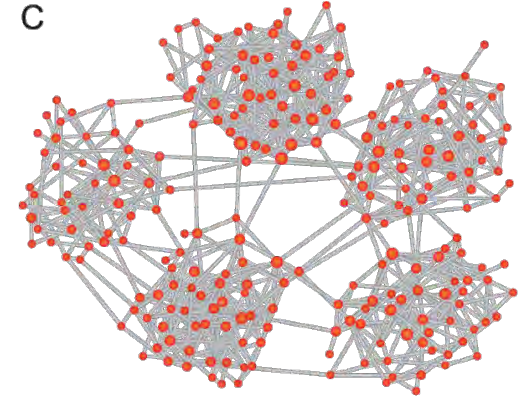
Scale-Free (SF) networks



Few nodes connected to many others (hubs)

Degree distribution follows a power law

Small-World (SW) networks



Between totally regular and random

Highly clustered but short path length

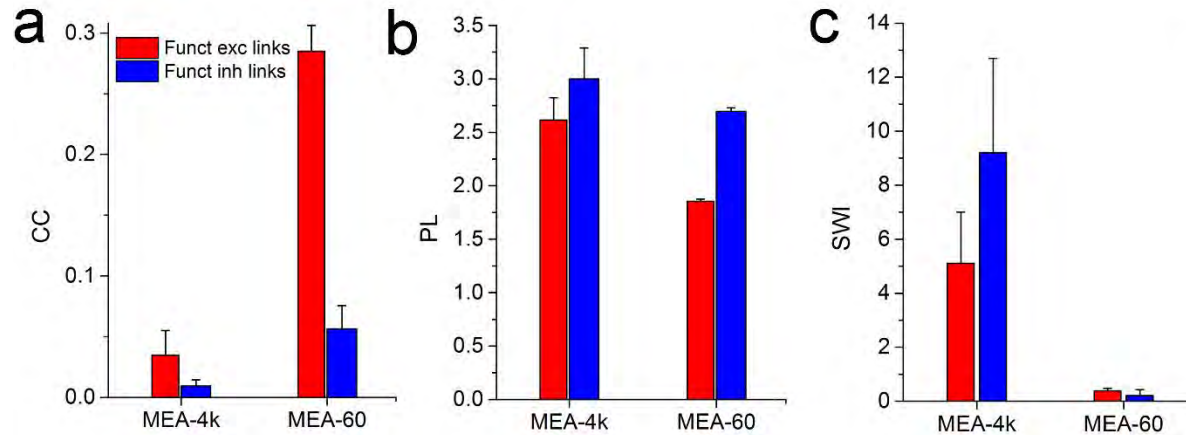
Erdos P., Renyi A. Publicationes Mathematicae, Vol. 6, pp. 290-297, 1959

Barabási A-L., Albert R. Science, Vol. 286, pp.509-512, 1999.

Watts D.J., Strogats S.H. Nature, Vol. 393, pp.440-442, 1998.

B+C: It is hypothesized to reflect an optimal configuration associated with rapid synchronization and information transfer

Network topology: MEA-60 and MEA-4k



Small-world topological properties found in large-scale networks

Scale-free networks

Rich-club: privileged sub-networks

Modulation of network dynamics

Modulation by chemical compounds: specific for receptors but difficult for a spatially confined delivery

Modulation by direct electrical stimulation: unspecific but spatially confined (you need an electrode properly placed)

What about **remote non invasive** neuro modulation?

Optogenetics and optical stimulation could be a partial answer.

Pros: specificity

Cons: still invasive; it implies a genetic modification of the cells...

Engineered networks with piezo-electric nanoparticles

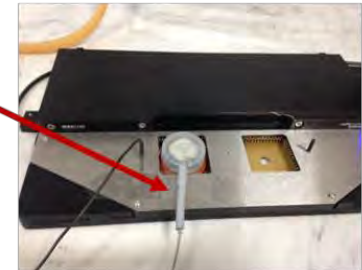
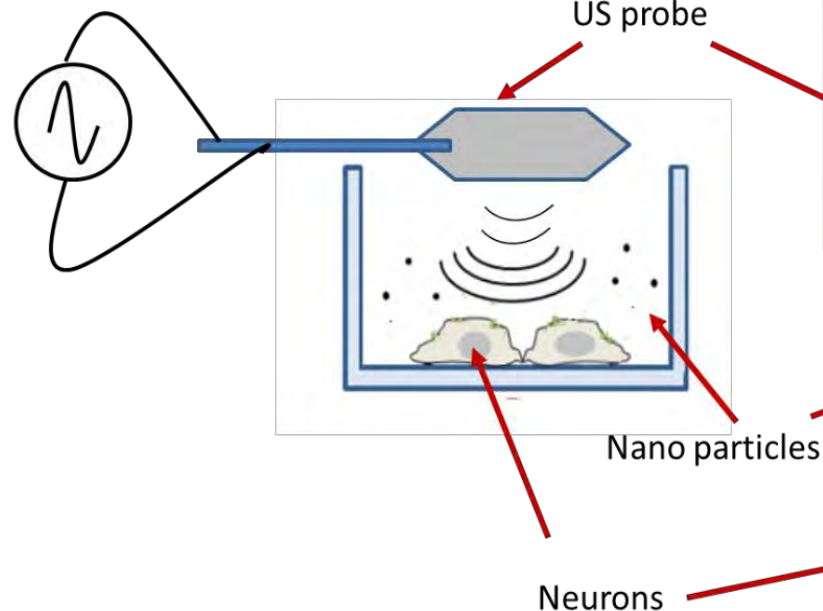


Camilo Rojas, PhD

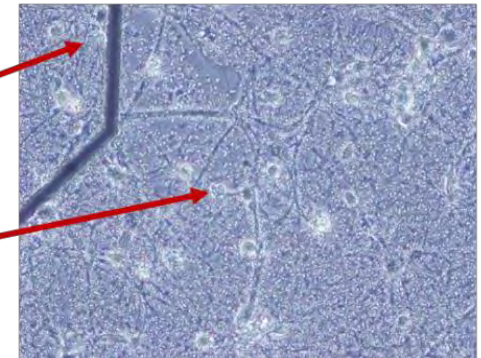
Barium titanate
nanoparticles
BTNP

ultrasound induced stimulation

Ultrasound source



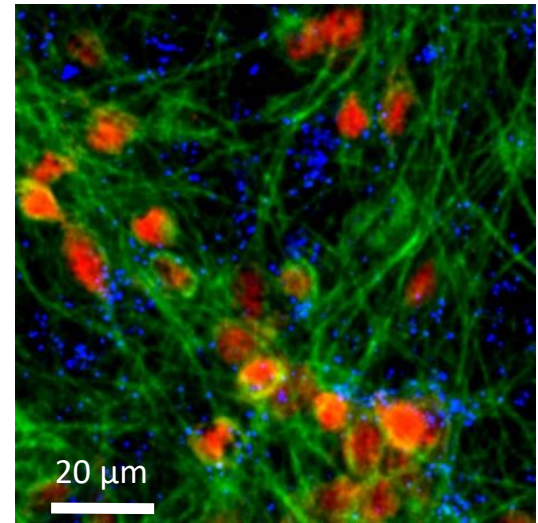
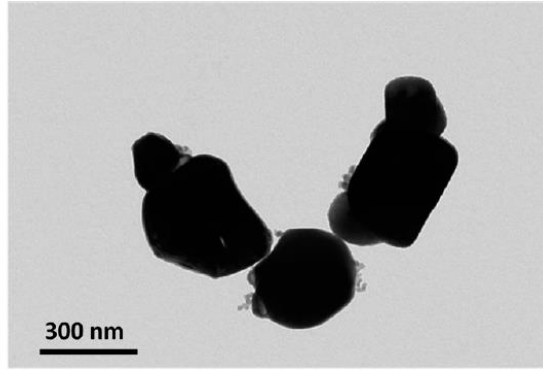
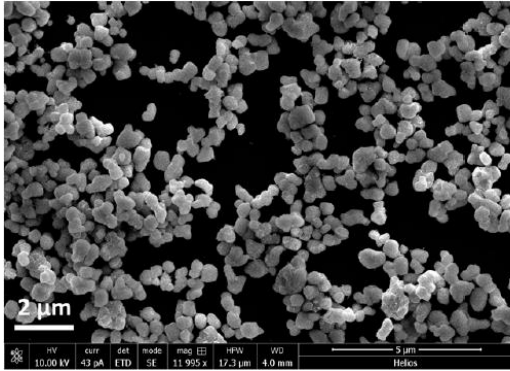
MEA2100, Multichannel Systems



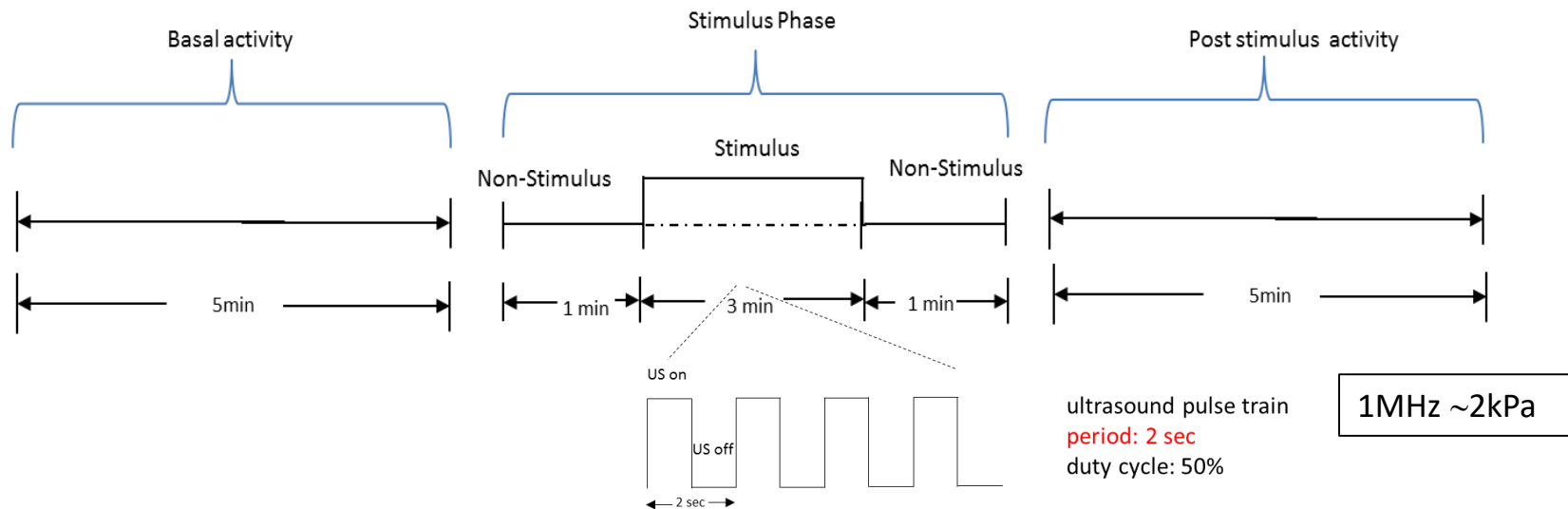
Gianni Ciofani



BaTiO₃ Nanoparticles



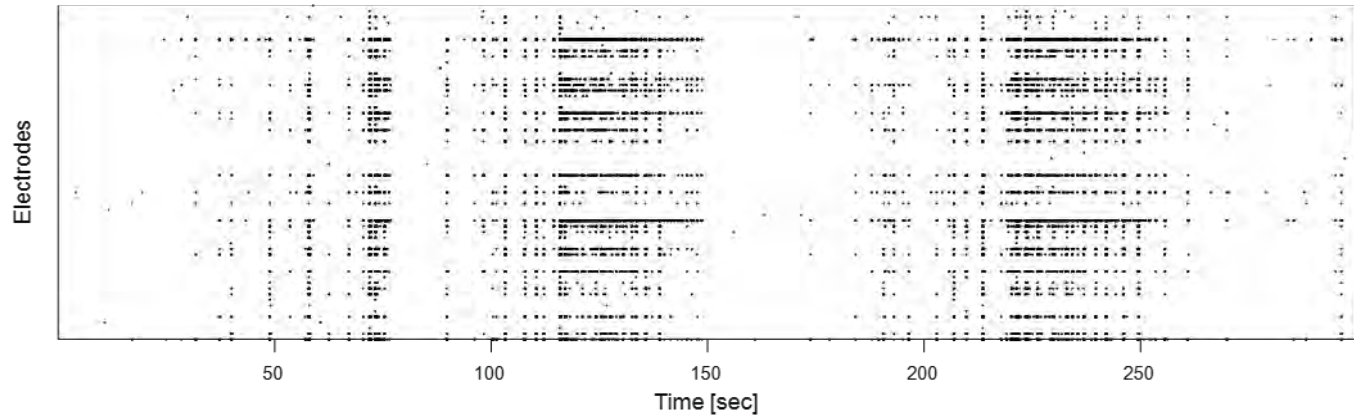
- wrapped in Arabic gum
- hydrodynamic size: 479.0 ± 145.3 nm (by DLS)
- biocompatible
- commercially available $\begin{cases} \text{with tetragonal crystalline phase (perovskite-like)} \rightarrow \text{piezoelectric} \\ \text{with cubic crystalline phase} \rightarrow \text{non-piezoelectric} \end{cases}$



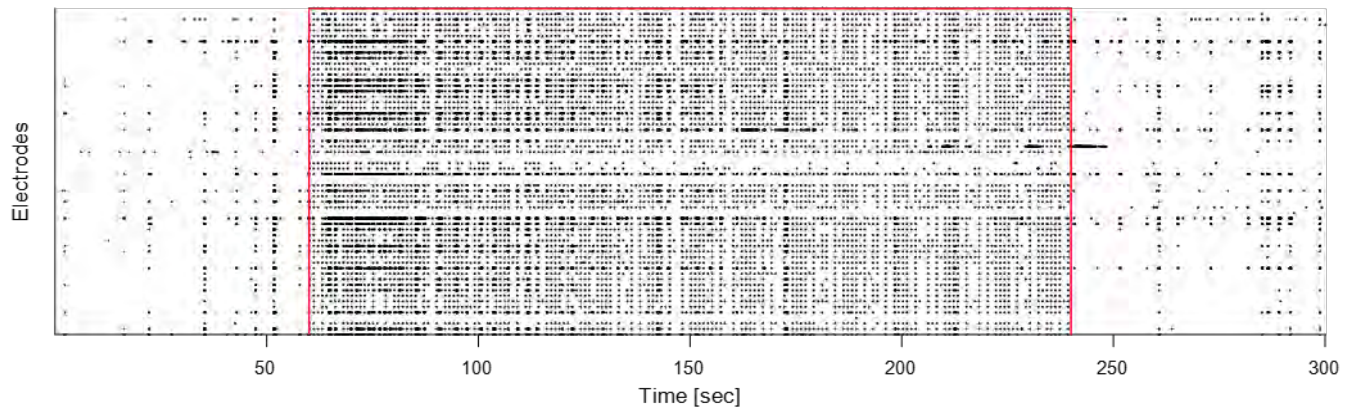
Acoustic stimulation: excitation

BTNP

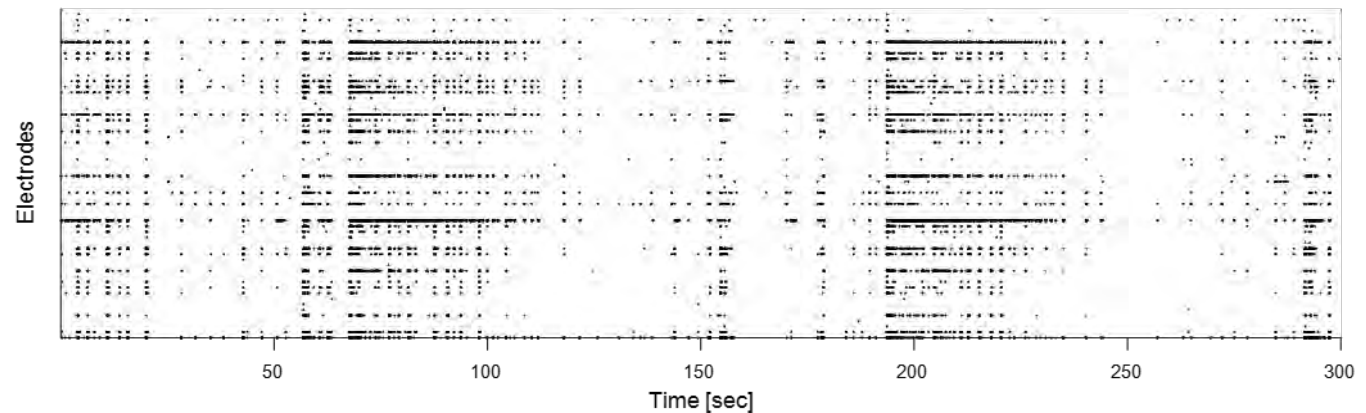
Base



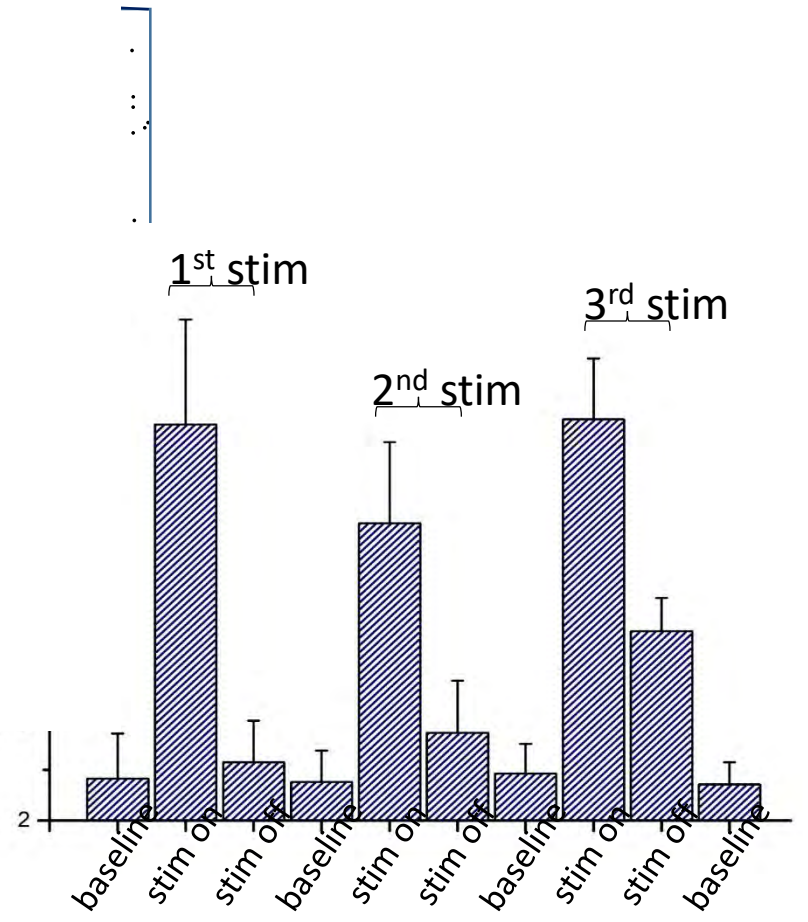
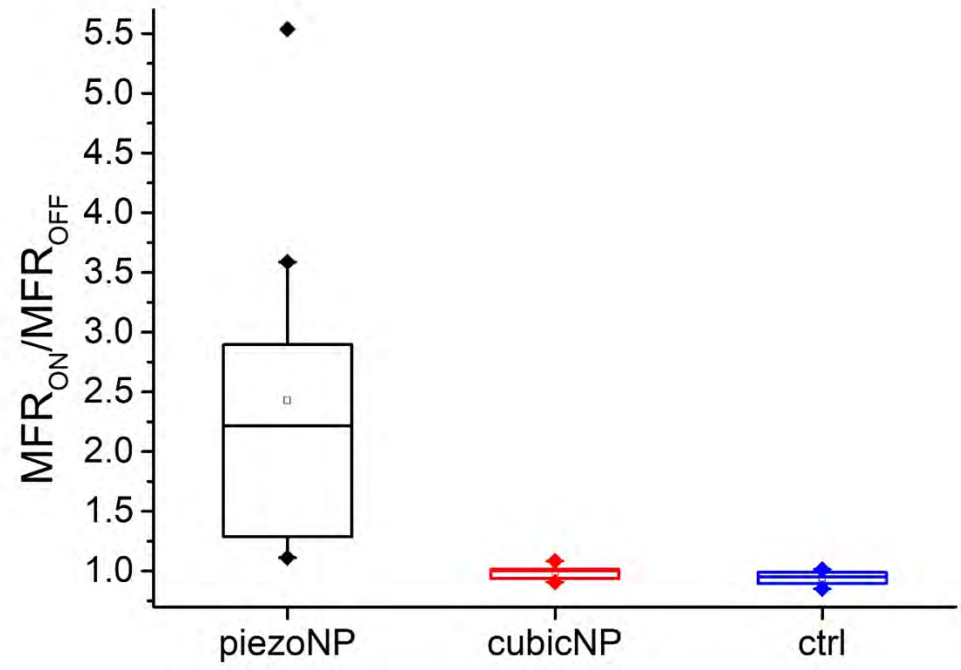
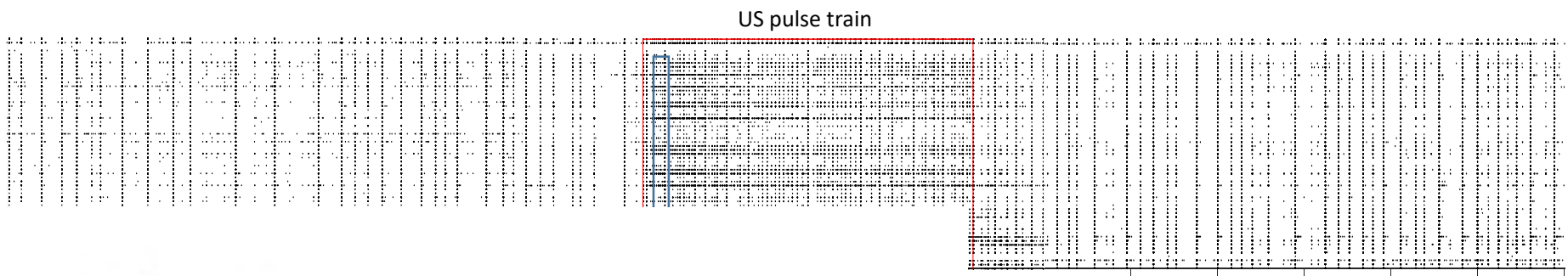
Stim



Post



Acoustic stimulation: excitation



US stimulation mediated by piezoelectric nanoparticles induces an excitatory response in cultured neural networks

Which mechanism?

A. Marino *et al.* *ACS Nano* Vol.9, 7678 (2015)

mechanical deformation



electro-elastic model of BTNP

local change in electric potential



increased open probability of voltage-gated channels



action potential

Engineered networks with Gold Nano Rods



Andrea Andolfi, MS

Gold Nano Rods GNR

When gold particles are synthesized at the nanoscale they improve their surface plasmon resonance, acquiring very interesting plasmonic properties. Thanks to these properties, gold nanoparticles find numerous applications in different fields, such as:

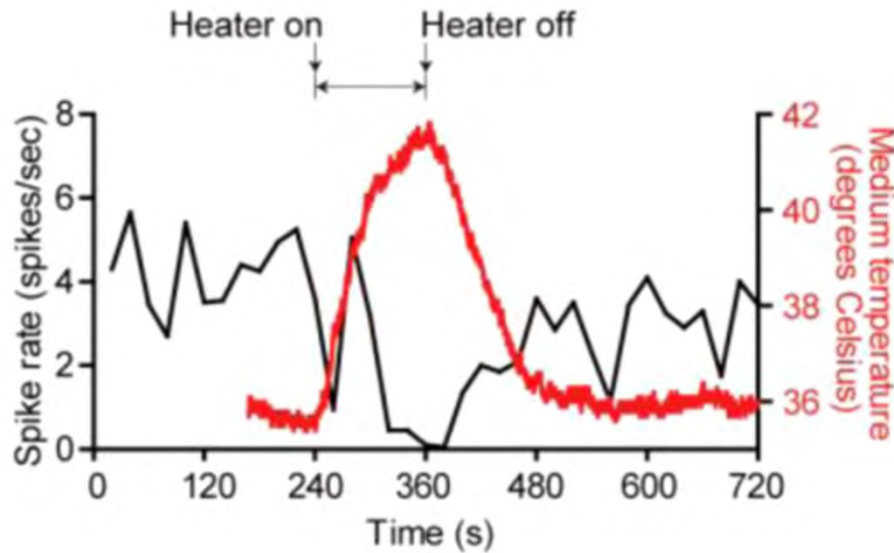
- Cancer therapy
- Biomedical sensing and imaging
- Drug delivery
- Nanophotonics
- **Neural activity modulation**

Yoonkey Nam



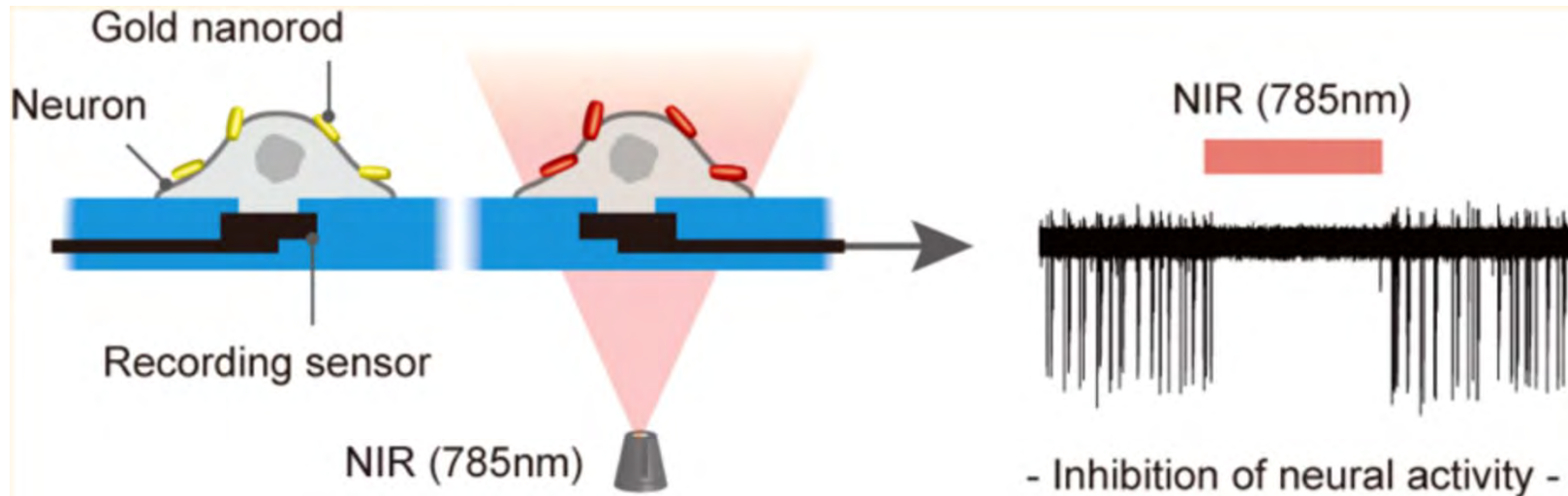
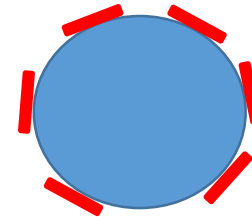
Korean Advanced Institute of
Science and Technology

Engineered networks with GNRs: photo thermal inhibition

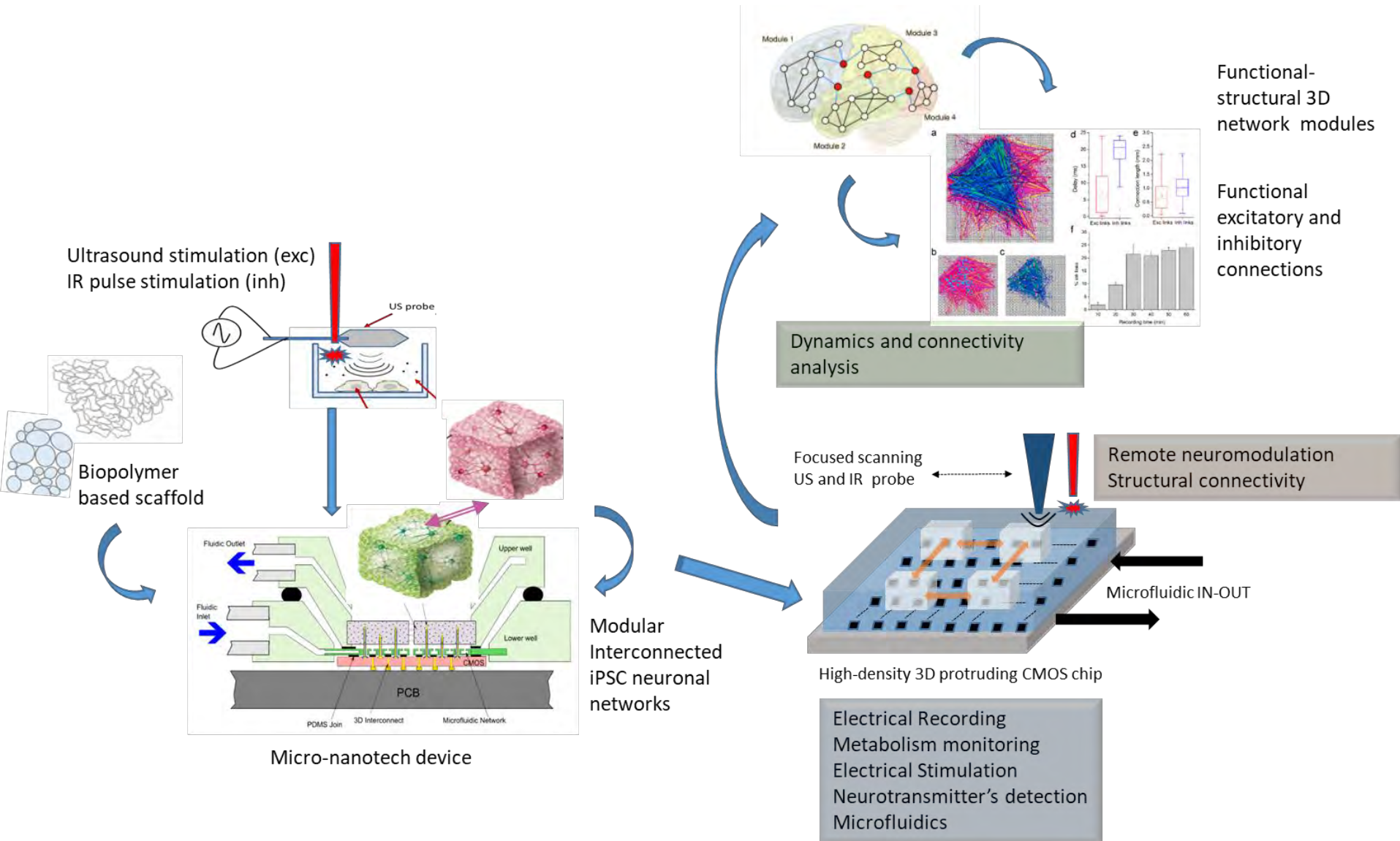


macroscale

Functionalized
microbeads



Engineered microsystems: brain-on-a-chip



Summary and conclusions

Tools and technologies for analyzing engineered model systems: e.g., high-density large scale MEA devices

Reliable analysis methods to infer connectivity. Ground truth problem, in silico models, in vitro models. Connectivity methods are at the basis to infer topology. A large number of nodes is needed...

Further engineered neuronal systems with nanoparticles for neural activity modulation:

Piezo nano-particles for stimulation

Gold nano-rods for inhibition

In vitro 3D models for brain-on-a-chip applications, towards engineered brain organoids and patient specific medicine

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Laura Pastorino (biomaterials)

Pasqualina Farisello (cell biology)

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Thanks for your attention!

School of NeuroEngineering "Massimo Grattarola"

